# International Union of Pharmacology. LIII. Nomenclature and Molecular Relationships of Voltage-Gated Potassium Channels

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### Introduction

Potassium-selective channels are the largest and most diverse group of ion channels, represented by some 70 known loci in the mammalian genome. The first cloned potassium channel gene was the Drosophila voltagegated shaker channel, and this was rapidly followed by the identification of other voltage- and ligand-gated potassium channel genes in flies, mammals, and many other organisms. The voltage-gated K<sub>v</sub> channels, in turn, form the largest family of some 40 genes among the group of human potassium channels, which also includes the  $\mathrm{Ca^{2+}}$ -activated ( $\mathrm{K_{Ca}}$ ), inward-rectifying ( $\mathrm{K_{IR}}$ ), and two-pore (K<sub>2P</sub>) families described in the following articles of this compendium. K<sub>v</sub> and K<sub>Ca</sub> channels together constitute the six/seven-transmembrane group of potassium-selective channels, made up of subunits containing six or seven membrane-spanning domains, including the positively charged S4 segment, which confers on some of these channels their voltage sensitivity.

Table 1 lists the International Union of Pharmacology (IUPHAR¹) names assigned to the members of the  $K_{\rm v}$  family of channels, as well as the gene names established by the HUGO Gene Nomenclature Committee (HGNC). Two new sequences,  $K_{\rm v}6.4$  and  $K_{\rm v}8.2$ , have been added to this list since the earlier edition of this compendium. Figures 1 and 2 show two phylogenetic tree reconstructions, one for the  $K_{\rm v}1$ –9 families and the other for the  $K_{\rm v}10$ –12 families, based on amino acid sequence alignments of the entire hydrophobic core of the proteins.

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<sup>1</sup> Abbreviations: IUPHAR, International Union of Pharmacology; HGNC, HUGO Gene Nomenclature Committee.

 $egin{array}{c} ext{TABLE 1} \ ext{$K_v$ channel families} \end{array}$ 

Gene names shown are those assigned by the IUPHAR (Catterall et al., 2002) and HGNC (http://www.gene.ucl.ac.uk) in addition to some other commonly used names.

mes.					
IUPHAR	HGNC	Other			
$K_v 1.1$ $K_v 1.2$ $K_v 1.3$ $K_v 1.4$ $K_v 1.5$ $K_v 1.6$ $K_v 1.7$ $K_v 1.8$	KCNA1 KCNA2 KCNA3 KCNA4 KCNA5 KCNA6 KCNA7	Shaker-related family			
$K_v2.1 \ K_v2.2$	KCNB1 KCNB2	Shab-related family			
$K_v 3.1 \ K_v 3.2 \ K_v 3.3 \ K_v 3.4$	KCNC1 KCNC2 KCNC3 KCNC4	Shaw-related family			
$K_v4.1 \ K_v4.2 \ K_v4.3$	KCND1 KCND2 KCND3	Shal-related family			
$K_v 5.1$	KCNF1	Modifier			
$K_v 6.1 \ K_v 6.2 \ K_v 6.3 \ K_v 6.4$	KCNG1 KCNG2 KCNG3 KCNG4	Modifiers			
$K_{v}7.1 \ K_{v}7.2 \ K_{v}7.3 \ K_{v}7.4 \ K_{v}7.5$	KCNQ1 KCNQ2 KCNQ3 KCNQ4 KCNQ5	KVLQT KQT2			
$K_v 8.1 \ K_v 8.2$	KCNV1 KCNV2	Modifiers			
$K_v 9.1 \ K_v 9.2 \ K_v 9.3$	KCNS1 KCNS2 KCNS3	Modifiers			
$K_v 10.1 \ K_v 10.2$	KCNH1 KCNH5	eag1 eag2			
$K_v 11.1 \ K_v 11.2 \ K_v 11.3 \ K_v 12.1 \ K_v 12.2 \ K_v 12.3$	KCNH2 KCNH6 KCNH7 KCNH8 KCNH3 KCNH4	erg1 erg2 erg3 elk1, elk3 elk2 elk1			

### Phylogenetic Tree, Kv1-9 Families

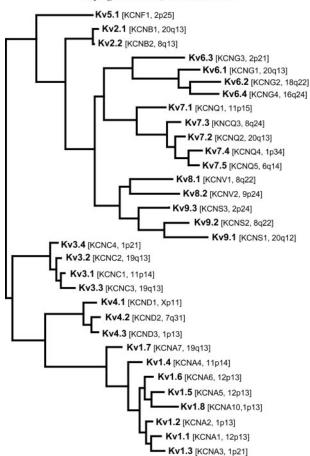


Fig. 1. Phylogenetic tree for the  $K_v1-9$  families. Amino acid sequence alignments of the human channel K<sub>v</sub> proteins were created using CLUSTALW, and analysis by maximum parsimony using PAUP\* resulted in unrooted trees comprising the K-1-K-6 and K-8-K-9 families that appeared in the previous edition of this compendium. Sequences of K,7.1-7.5, K,6.4, and K,8.2 were added to the existing alignment, and these new sequences were incorporated into the existing tree topology by use of a combination of maximum parsimony and neighbor-joining analvsis. Only the hydrophobic cores (S1–S6) were used for analysis. The IUPHAR and HGNC names are shown together with the genes' chromosomal localization and other commonly used names.

K<sub>v</sub> channels form an exceedingly diverse group, much more so than one would predict simply based on the number of distinct genes that encode them. This diversity arises from several factors. 1) Heteromultimerization. Each K, gene encodes a peptide subunit, four of which are required to form a functional channel. K, channels may be homotetramers but may also be heterotetramers formed between different subunits within the same family (in the case of the K<sub>v</sub>1, K<sub>v</sub>7, and K<sub>v</sub>10 families), and these diverse heterotetramers express properties that may be considerably different from those of any of the homotetramers. 2) "Modifier" subunits. Four of the K, families (K, 5, 6, 8, and 9) encode subunits that act as modifiers. Although these do not produce functional channels on their own, they form heterotetramers with K<sub>2</sub> family subunits, increasing the functional diversity within this family. 3) Accessory proteins. A variety of other peptides has also been shown to

### Phylogenetic Tree, Kv10-12 Families

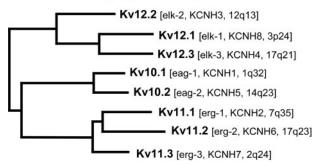


Fig. 2. Phylogenetic tree for the  $K_{\nu}10{-}12$  families. This unrooted tree was created as described in Fig. 1 and appeared in the previous edition of this compendium. The IUPHAR and HGNC names are shown together with the genes' chromosomal localization and other commonly used names.

associate with K<sub>v</sub> tetramers and modify their properties, including several  $\beta$  subunits (which associate with K.,1 and K,2 channels), KCHIP1 (K,4), calmodulin (K,10), and minK (K<sub>v</sub>11), as well as many others identified in the tables that follow the text of this article. 4) Alternate mRNA splicing. A number of K<sub>v</sub> channel genes are known to contain intronless coding regions, including all of the K<sub>2</sub>1 family genes (with the sole exception of K<sub>2</sub>1.7) and K<sub>v</sub>9.3. Although alternate splicing of noncoding exons may be important in regulating the expression of these channels, one gene can produce only a single kind of protein subunit. However, various members of the K<sub>v</sub>3, 4, 6, 7, 9, 10, and 11 gene families have coding regions made up of several exons that are alternately spliced, providing yet another significant source of K, channel functional diversity. 5) Post-translational modification. Many K, channels can be post-translationally modified by phosphorylation (Jerng et al., 2004), ubiquitinvlation (Henke et al., 2004), and palmitoylation (Gubitosi-Klug et al., 2005), which in turn modifies channel function.

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Our current understanding of the roles of this family of channels is catalogued in Tables 2 through 41, including recent developments in the pharmacology, regulation of expression, and disease associations of its various members (Misonou and Trimmer, 2004; Norton et al., 2004; Wua and Dworetzky, 2005).

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### TABLE 2 $K_V 1.1$ channels

 $K_V 1.1^{1-6}$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names HuK (I), MBK1, MK1, RCK1, RBK1, HBK1

Human: 494 aa, NM\_000217, chr. 12p13.3, 7.8 KCNA1, GeneID: 3736, PMID: 134929735 Molecular information

> Mouse: 495aa, NM\_010595, chr. 6 Rat: 495aa, NM\_173095, chr. 4q42

 $K_V\beta_1$ ,  $K_V\beta_2$ , PSD95, synapse-associated protein 97 (SAP97), SNAP25<sup>9-19</sup> Associated subunits

Functional assays Voltage-clamp

Current Voltage-gated potassium channel in neurons and skeletal muscle

Conductance

Ion selectivity  $K^{+}(1) > Rb^{+}(0.8) > NH_{4}^{+}(0.1)$ 

 $\begin{array}{l} V_{\rm a} = -32~{\rm mV}; \, k_{\rm a} = 8.5~{\rm mV}; \, \tau_{\rm n} = 5~{\rm ms} \; (-32~{\rm mV})^{20,21} \\ V_{\rm h} = -51~{\rm mV}; \, k_{\rm h} = 3~{\rm mV}; \, \tau_{\rm h} = 11~{\rm s} \; (40~{\rm mV})^{20,21} \end{array}$ Activation Inactivation

Activators Gating inhibitors None

Blockers Tetraethyammonium (0.3 mM), DTX (20 nM), DTX-K, ShK (16 pM), 10-N-methylcarbamoyl-3,7-

bis(dimethylamino)phenothiazine (490 nM), 4-aminopyridine (290  $\mu$ M), capsaicin (29  $\mu$ M), resiniferatoxin (9 µM), flecainide (209 µM), nifedipine (96 µM), diltiazem (144 µM), kaliotoxin

(41 nM), hongotoxin-1, margatoxin<sup>20,22-24</sup>

<sup>125</sup>I-DTX, <sup>125</sup>I-BgK<sup>25,26</sup> Radioligands

Brain, heart, retina, skeletal muscle, islets<sup>27–31</sup> Channel distribution

Maintaining membrane potential, modulating electrical excitability in neurons and muscle Physiological functions

Mutations and pathophysiology Episodic ataxia/myokymia syndrome type 18,32-34

Pharmacological significance

Comments  $K_V1.1$  can coassemble with others in the  $K_V1$  family members in heteromultimers, but not with members of other K<sub>V</sub> families; introlless coding region; mammalian Shaker-related family

aa, amino acids; chr., chromosome; DTX, dendrotoxin; ShK, Stychodactyla helianthus toxin; BgK, Bundosoma granulifera toxin.

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### TABLE 3 $K_V 1.2$ channels

Channel name  $K_v 1.2$ 

Description Voltage-gated potassium channel, delayed rectifier

HuK (IV), MK2, BK2, RCK5, RAK, BGK5, XSha2, NGK1, HBK5<sup>1-8</sup> Other names

Molecular information Human: 499aa, NM\_004974, chr. 1p13, KCNA2, GeneID: 3737, PMID: 225128333

> Mouse: 499aa, NM\_008417, chr. 3 Rat: 499aa, NM\_012970 chr. 2q34

K<sub>V</sub>β<sub>1</sub>, K<sub>V</sub>β<sub>2</sub>, PSD95, synapse-associated protein 97 (SAP97), SNAP25, Caspr2, RhoA<sup>9-17</sup> Associated subunits

Functional assays Voltage-clamp Current Delayed rectifier  $14-18 pS^{18}$ Conductance Ion selectivity K<sup>+</sup>-selective

Voltage-dependent,  $V_{\rm a}$  between 5 and 27 mV;  $k_{\rm a}$  = 13 mV;  $\tau_{\rm n}$  = 6 ms (60 mV)<sup>6,18</sup> Activation

 $V_{\rm h}$  between -33 and -15 mV;  $k_{\rm h} \sim 8$  mV<sup>6,18</sup> Inactivation

Activators None Gating inhibitors None

Blockers 4-Aminopyridine (590  $\mu$ M), capsaicin (45  $\mu$ M), resiniferatoxin (31  $\mu$ M), flecainide (217  $\mu$ M),

nifedipine (18  $\mu$ M), diltiazem (187  $\mu$ M), 10-N-methylcarbamoyl-3,7-

bis(dimethylamino)phenothiazine (0.44 µM), DTX (17 nM), charybdotoxin (14 nM), margatoxin, natrexone (2 nM), tetraethyammonium (560 mM), H37 (18  $\mu$ M), picrotoxin-K $\alpha$  (32 pM), OsK2

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(97 nM), BgK (25 nM), HgTx (pM), anandamide (2.7 μM)<sup>18–23</sup>

 $^{125}$ I-DTX,  $^{125}$ I-HgTX1-A19Y/Y37F $^{22}$ Radioligands

Channel distribution Brain (pons, medulla, cerebellum, inferior colliculus > hippocampus, thalamus, cerebral cortex,

superior colliculus > midbrain, corpus striatum, olfactory bulb; neurons associated with

mechanoreception and proprioception), spinal cord, Schwann cells, atrium, ventricle, islet, retina,

smooth muscle, PC12 cells 1-8,24-30

Physiological functions Mutations and pathophysiology

Pharmacological significance

Comments

Maintaining membrane potential, modulating electrical excitability in neurons and muscle Not established

Not established

Delayed rectifier potassium channel; can coassemble with other K<sub>V</sub>1 family members in

heteromultimers but not with members of other  $K_V$  families  $^{19,22,25,29,31}$ ; intronless coding region  $^5$ ; T1 domain in N terminus required for multimerization<sup>32</sup>; mammalian Shaker-related family

aa, amino acids; chr., chromosome; DTX, dendrotoxin; HgTX, hongotoxin.

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### TABLE 4 $K_V 1.3$ channels

 $K_v 1.3^{1-8}$ Channel name Description Voltage-gated potassium channel, delayed rectifier Other names MK3, MBK3, RCK3, hPCN3, HuK (III), HLK3, RGK5, KV3, HGK5, n-channel Human: 523aa, NM\_002232, chr. 1p13.3,  $^{7,9}$  KCNA3, GeneID: 3738, PMID: 2251283 $^4$ Molecular information Mouse: 528aa, NM\_008418, chr. 3 Rat: 525aa, NM\_019270, chr. 2q34 Associated subunits  $K_V\beta$ , hDlg,  $\beta_1$  integrin, KChaP<sup>10-12</sup> Functional assays Voltage-clamp Type N voltage-gated potassium channel in lymphocytes<sup>3,4</sup> Current  $13pS^4$ Conductance  $K^+$  (1)  $> Rb^+$  (0.77)  $> NH_4^+$  (0.1)  $> Cs^+$  (0.02)  $> Na^+$  (<0.01)<sup>13</sup> Ion selectivity Voltage,  $V_{\rm a}=-35$  mV;  $k_{\rm a}=6$  mV;  $\tau_{\rm n}=3$  ms at 40 mV<sup>4,13</sup> Activation Inactivation C-type inactivation,  $V_{\rm h} = -63$  mV;  $k_{\rm h} = 7.7$  mV;  $\tau_{\rm h} = 250$  ms (40 mV) $^4$ Activators None None Gating inhibitors Blockers 4-Aminopyridine (195 μM), tetraethyammonium (10 mM), charybdotoxin (3 nM), naltrexone (1 nM), MgTX (110 pM), kaliotoxin (650 pM), AgTX2 (200 pM), Pi1 (11 nM), Pi2 (50 pM), Pi3 (500 pM),  $HsTx1~(12~pM),~ShK~(11~pM),~BgK~(39~nM),~ShK-Dap22~(52~pM),~quinine~(14~\mu\text{M}),~diltiazem$ (60 μM), verapamil (6 μM), CP339818 (150 nM), UK78282 (200 nM), correolide (90 nM), sulfamid-

benzamidoindane (100 nM), capsaicin (26 μM), resiniferatoxin (3 μM), nifedipine (5 μM), H37  $(23~\mu M)^{14,15}$ <sup>125</sup>I-HgTx1-A19Y/Y37F mutant (0.1–0.25 pM); <sup>125</sup>I-MgTx (0.3 pM)<sup>16,17</sup>

Brain (inferior colliculus > olfactory bulb, pons/medulla > midbrain, superior colliculus, corpus striatum, hippocampus, cerebral cortex), lung, islets, thymus, spleen, lymph node, fibroblasts, B lymphocytes, T lymphocytes, pre-B cells, tonsils, macrophages, microglia, oligodendrocytes, osteoclasts, platelets, testis  $^{1-8,18-21}$ 

Regulation of membrane potential and calcium signaling in lymphocytes and oligodendrocytes 14,21-23 Not established

Therapeutic target for immunosuppressants; K<sub>V</sub>1.3 inhibitors suppress T-cell activation in vitro and delayed type hypersensitivity in vivo and have proven effective for multiple sclerosis in an animal $^{24,25}$ ;  $K_v I.3$  expression is dramatically and exclusively increased in effector memory T cells Can coassemble with other K<sub>V</sub>1 family members in heteromultimers but not with members of other K<sub>V</sub> families; introlless coding region; mammalian Shaker-related family

Comments

Radioligands

Channel distribution

Physiological functions Mutations and pathophysiology

Pharmacological significance

aa, amino acids; chr., chromosome; MgTX, margatoxin; HgTX, hongotoxin; CP339818, N-[1-(phenylmethyl)-4(1H)-quinolinylidene]-1-pentamine monohydrochloride; UK78282, 4-[(diphenylmethoxy)methyl]-1-[3-(4-methoxyphenyl)propyl]-piperidine.

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### TABLE 5 $K_{\rm V}1.4$ channels

 $K_V 1.4^{1-7}$ Channel name

Description Voltage-gated potassium channel, A-type, fast-inactivating Other names HuK (II), hPCN2, HK1, RCK4, RHK1, RK4, RK8, MK4

Molecular information Human: 653aa, NM\_002233, chr. 11p14.3-15.2, KCNA4, GeneID: 3739, PMID: 2263489<sup>32</sup>

> Mouse: 654aa, NM\_021275, chr. 2 Rat: 654aa, NM\_012971, chr. 3q33

 $K_V\beta$ , PSD95, synapse-associated protein 97 (SAP97), SAP90,  $\alpha$ -actinin-2, KChaP,  $\sigma$  receptor<sup>8–18</sup> Associated subunits

Functional assays Voltage-clamp

Current  $K_V 1.4/K_V 1.2$  heteromultimers may underlie the presynaptic A-type K<sup>+</sup> channel<sup>19</sup>

Conductance  $5pS^1$ 

K<sup>+</sup>-selective (50 times more selective for K<sup>+</sup> than Na<sup>+</sup>)<sup>20</sup> Ion selectivity

Voltage,  $V_a = -22 \text{ mV}^1$ ;  $-34 \text{ mV}^{20}$ ;  $K_a = 5^{21}$ Activation

N-type inactivation,  $V_h = -62 \text{ mV}^{20}$ ;  $\tau_h = 47 \text{ ms } (0 \text{ mV})^{20}$ Inactivation

CaMKII/calcineurin regulation through phosphorylation/dephosphorylation makes inactivation Activators

Ca<sup>2+</sup>-dependent<sup>22</sup>

Gating inhibitors

4-Aminopyridine (13  $\mu$ M), <sup>1</sup> tetraethyammonium (>100 mM), <sup>3</sup> UK78282 (170 nM), <sup>23</sup> riluzole Blockers

 $(70 \mu M)^{24}$  quinidine  $(10 \mu M-1 mM)^{25}$  nicardipine  $(0.8 \mu M)^{26}$ 

Radioligands None

Channel distribution Brain (olfactory bulb, corpus striatum > hippocampus, superior and inferior colliculus > cerebral

cortex, midbrain basal ganglia > pons/medulla), lung-carcinoid, skeletal muscle, heart, pancreatic

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 $islet^{1,6,27-29}$ 

Neuronal afterhypolarization Physiological functions

Mutations and pathophysiology K<sub>V</sub>1.4 expression increases in rat ventricular myocytes after myocardial infarction and induction of

diabetes 30,31 Not established

Pharmacological significance

Comments

Can coassemble with other K, 1 family members in heteromultimers but not with members of other  $K_v$  families; intronless coding region; mouse  $K_v$ 1.4 mRNA contains an internal ribosome entry site

in its 5'-noncoding region and may be translated by cap-independent mechanisms<sup>33,34</sup>;

mammalian Shaker-related family

aa, amino acids; chr., chromosome

<sup>1.</sup> Stuhmer W, Ruppersberg JP, Schroter KH, Sakmann B, Stocker M, Giese KP, Perschke A, Baumann A, and Pongs O (1989) Molecular basis of functional diversity of voltage-gated potassium channels in mammalian brain. EMBO J 8:3235-3244.

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### TABLE 6 $K_V 1.5$ channels

 $K_{\rm V}1.5$ Channel name

Voltage-gated potassium channel, delayed rectifier Description HpCN1, HK2, HCK1, KV1, fHK, RK3, RMK2, HuK (II)<sup>1-8</sup> Other names

Human: 613aa, NM\_002234, chr. 12p13.3,8-10 KCNA5, GeneID: 3741, PMID: 19863823 Molecular information

> Mouse: 602aa, NM\_002234, chr. 6 Rat: 602aa, NM\_012972, chr. 4q42-44

 $K_V\beta_1$ ,  $K_V\beta_2$ , KCNA3B, Src tyrosine kinase, fyn, KChaP,  $\alpha$ -actinin-2, caveolin, synapse-associated Associated subunits

protein 97  $(SAP97)^{11-21}$ 

Voltage-clamp Functional assays

Ultrarapid-activating K<sup>+</sup> current in heart (IK<sub>ur</sub>)<sup>22,23</sup> Current

Not established

Conductance Ion selectivity

Activation

Voltage,  $V_{\rm a}=-14$  mV;  $k_{\rm a}=6$ –12 mV<sup>22,24</sup>  $V_{\rm h}=-25$  to -10 mV;  $k_{\rm h}=3$ –5 mV;  $\tau_{\rm h1}=460$  ms;  $\tau_{\rm h2}=5$  s (40 mV)<sup>22,24</sup> Inactivation

Activators Gating inhibitors None

Blockers S9947 (420 nM), 4-aminopyridine (270  $\mu$ M),capsaicin (23  $\mu$ M), resiniferatoxin (26  $\mu$ M), flecainide

 $(101~\mu\text{M})$ , nifedipine  $(81~\mu\text{M})$ , diltiazem  $(115~\mu\text{M})$ , tetraethyammonium (330~mM), clofilium inside

(140 nM), bupivacaine (4.1  $\mu$ M), propafenone (4.4  $\mu$ M),  $^{24-26}$  quinidine (0.6  $\mu$ M) $^{27}$ 

Radioligands None

Channel distribution Aorta, colon, kidney, pooled colon, kidney, stomach, smooth muscle, whole embryo, hippocampus and

cortex (oligodendrocytes, microglia, Schwann cells), pituitary, pulmonary artery<sup>1-7,28-33</sup>

Physiological functions  $K_{\rm V}1.5$  has properties similar to the ultrarapidly activating  $IK_{\rm ur}$  current in the heart, and antisense-

targeting  $K_V 1.5$  suppresses  $IK_{ur}$  currents almost  $50\%^{22,23}$ ; maintains membrane potential that

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modulates electrical excitability in neurons

Mutations and pathophysiology Pharmacological significance

Potential use in management of atrial fibrillation via blockade of IK<sub>ur</sub> <sup>34,35</sup>

Comments Can coassemble with other K<sub>V</sub>1 family members in heteromultimers but not with members of other K<sub>V</sub> families; intronless coding region; mammalian *Shaker*-related family.

aa, amino acids; chr., chromosome,

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### TABLE 7 $K_V 1.6$ channels

Channel name  $K_{v}1.6^{1-5}$ 

Description Voltage-gated potassium channel, delayed rectifier

Other names HBK2, MK1.6, RCK2, KV2

Molecular information Human: 528aa, NM\_002235, chr. 12p13.3,6 KCNA6, GeneID: 3742, PMID:23473051

Mouse: 529aa, NM\_013568, chr. 6

Rat: 530 aa, XM\_575671 (predicted), chr. 4q42

 $K_V\beta_1$ ,  $K_V\beta_2$ ,  $^{7,8}$   $Caspr2^{18}$ Associated subunits Functional assays Voltage-clamp

Current Delayed rectifier Conductance  $9pS^1$ 

Ion selectivity K<sup>+</sup>-selective

 $V_{\rm a} = -20 \text{ mV}; k_{\rm a} = 8 \text{ mV}^1$ Activation  $K_{\rm h} = -43^2; \, \tau_{\rm h} = > 3 \, \, {
m s}^1$ Inactivation

Activators None Gating inhibitors

α-Dendrotoxin (20 nM), 10-N-methylcarbamoyl-3,7-bis(dimethylamino)phenothiazine (10 nM, 1 Blockers

200 nM<sup>3</sup>), 4-aminopyridine (1.5 mM), <sup>1,3</sup> tetraethyammonium (7 mM), <sup>1,3</sup> ShK (160 pM), <sup>9</sup> HgTX (9.6 pM), 10 BgK (W5Y/F6A/Y26F)11

Radioligands <sup>125</sup>I-BgK (W5Y/F6A/Y26F), <sup>11</sup> <sup>125</sup>I-HgTX

Channel distribution Brain, colon, germ cell, heart, lung, ovary, testis, astrocytes, pulmonary artery smooth muscle cells,

oligodendrocytes<sup>1,3-5,8,12-16</sup>

Physiological functions Regulator of membrane potential in neurons

Mutations and pathophysiology No K<sup>+</sup> channel clustering in optic nerves of hypomyelinating Shiverer mice

Pharmacological significance Not established

Comments Can coassemble with other K<sub>V</sub>1 family members in heteromultimers but not with members of other

K<sub>v</sub> families; introlless coding region; N terminus contains an N terminus inactivation prevention

(NIP) domain;17 mammalian Shaker-related family

aa, amino acids; chr., chromosome; HgTx, hongotoxin.

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TABLE 8  $K_V 1.7$  channels

 $K_{v}1.7^{1-3}$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names

Human: 456aa, NM\_031886, chr. 19q13.3<sup>1-3</sup>, KCNA7, GeneID: 3743, PMID: 11368907<sup>6</sup> Molecular information

Mouse: 532aa, NM\_010596, chr. 7

Rat: 457, XM\_344889 (predicted), chr. 1q22

Associated subunits None identified Functional assays Voltage-clamp

Current Possibly a component of IK<sub>ur</sub> in the heart<sup>3</sup>

Conductance  $\mathbf{K}^{+}$ Ion Selectivity

Voltage,  $V_a = -8 \text{ mV}; \tau_n = 6 \text{ ms} (30 \text{ mV})^3$ Activation

Very slow inactivation Inactivation

Activators None Gating inhibitors None

Blockers Flecainide (8  $\mu$ M), quinidine (15  $\mu$ M), verapamil (16  $\mu$ M), amiodarone (35  $\mu$ M), 4-aminopyridine

 $(150 \mu M)$ , tetraethyammonium  $(150 \text{ mM})^3$ 

Radioligands None

Channel distribution Placenta, amnion, islets (mouse), skeletal muscle, heart, pulmonary arteries<sup>4,5</sup> Physiological functions  $K_V 1.7$  has properties similar to the ultrarapidly activating  $IK_{ur}$  current in the heart<sup>3</sup>

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments Can coassemble with other  $K_V$ 1 family members in heteromultimers but not with members of other  $K_V$ 

families; only member of this family that has an intron in the coding region 1-3; mammalian Shaker-

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related family

aa, amino acids; chr., chromosome.

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### TABLE 9 $K_{\rm V}1.8$ channels

Channel name  $K_v 1.8$ 

Description Voltage-gated potassium channel, delayed rectifier

Other names K<sub>v</sub>1.10, Kcn1<sup>1</sup>

Human: 511aa, NM 005549, chr. 1p13.1, KCNA10, GeneID: 3744, PMID: 9177773<sup>1</sup> Molecular information

Mouse: 503aa, XM\_143471 (predicted), chr. 3 Rat: 511aa, XM\_227577 (predicted), chr. 2q34

Associated subunits KCNA4B Functional assays Voltage-clamp

Possibly a component of IK, in the heart<sup>2</sup> Current

 $10-12pS^{2}$ Conductance  $K^+/Na^+ > 70:1^2$ Ion selectivity

Activation  $V_a = 3.6 \text{ mV (oocytes)}; \tau_a = 18 \text{ ms at } +60 \text{ mV (oocytes)}^2$ 

 $\tau_h = 10 \text{ s}$ CGMP Inactivation Activators Gating inhibitors None

Blockers Barium (5 mM), tetraethyammonium (50 mM), 4-aminopyridine (1.5 mM), charybdotoxin (100 nM),

ketoconazole (500 nM), pimozide (300 nM), verapamil (45 μM)<sup>2</sup>

Radioligands

Channel distribution Kidney (cortex > medulla), brain, heart, skeletal muscle, adrenal gland 1-3,6

Physiological functions Regulation of membrane potential in renal proximal tubule

Mutations and pathophysiology None

Pharmacological significance Not established

Comments Can coassemble with other  $K_V 1$  family members in heteromultimers but not with members of other

K<sub>V</sub> families; intronless coding region; mammalian Shaker-related family

aa, amino acids; chr., chromosome.

4. Yao X, Liu Y, Tung F, and Desir GV (1996) Genomic structure and regulation of Kcn1, a cGMP-gated potassium channel. Am J Physiol 271:F37-F41.

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UniGene Cluster Hs0.306973; OMIM no. 176268.

<sup>1.</sup> Orias M, Bray-Ward P, Curran ME, Keating MT, and Desir GV (1997) Genomic localization of the human gene for KCNA10, a cGMP-activated K channel. Genomics 42:33–37. 2. Lang R, Lee G, Liu W, Tian S, Rafi H, OriasM, Segal AS, and Desir GV (2000) KCNA10: a novel ion channel functionally related to both voltage-gated potassium and CNG cation channels. Am J Physiol 278:F1013-F1021.

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TABLE 10  $K_{\rm v}2.1$  channels

 $K_{\rm V}2.1^{1-3}$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names hDRK1, DRK1

Human: 858aa, NM\_004975, chr. 20q13.2, 4.5 KCNB1, GeneID: 3745, PMID: 808172335 Molecular information

> Mouse: 857aa, NM\_008420, chr. 2 Rat: 853aa, NM\_013186, chr. 3q42

K<sub>V</sub>5.1, K<sub>V</sub>6.1-K<sub>V</sub>6.3, K<sub>V</sub>8.1, K<sub>V</sub>9.1-K<sub>V</sub>9.3, KChaP (binds to N terminus of K<sub>V</sub>2.1), Fyn SH2 Associated subunits

 ${\rm domain}^{6-15}$ 

Voltage-clamp Functional assays

K<sub>v</sub>2.1/K<sub>v</sub>9.3 (delayed rectifier in oxygen-sensitive pulmonary artery), delayed rectifier current in Current

hippocampal and globus pallidus neurons $^{16,17}$ 

8pS; on removal of  $K^+$ ,  $K_v2.1$  displays a large  $Na^+$  conductance that is inhibited by low Conductance

concentrations of  $K^{+2,12}$ 

Ion selectivity  $K^+ > Rb^+$ 

Activation Voltage,  $V_a = 12 \text{ mV}$ ;  $k_a = 3 \text{ mV}^3$ 

Inactivation Noninactivating Activators Linoleic acid19

Hanatoxin  $(42 \text{ nM})^{20,21}$ Gating inhibitors

Internal tetraethylammonium and tetrapentylammonium, internal Ba $^{2+}$  (13  $\mu$ M), external Ba $^{2+}$ Blockers

(30 mM), internal Mg<sup>2+</sup>, 4-AP (18 mM), halothane<sup>22–25</sup>

Radioligands None

Channel distribution Brain (cerebral cortex > hippocampus > cerebellum > olfactory bulb; restricted to neurons, where staining is present on dendrites and cell bodies but not on axons; Schwann cells), atria, ventricle,

skeletal muscle, retina, cochlea, eye, germ cell, lung, PC12 cells, pulmonary arteries,

Ser857Asn polymorphism in 0-3% in different ethnic populations<sup>5</sup>; two other single nucleotide

insulinomas  $^{1,3,9,14,16,17,26-33}$ 

Physiological functions Maintaining membrane potential and modulating electrical excitability in neurons and muscle 9,16,17

K<sub>v</sub>2.1 expression is reduced in chronic hypoxic pulmonary hypertension.<sup>30,32</sup> Mutations and pathophysiology

Pharmacological significance Comments

polymorphisms have been identified<sup>34</sup>; the 4-AP binding site is in the S6 inner vestibule.<sup>23</sup> Mammalian Shab-related family.

aa, amino acids; chr., chromosome; 4-AP, 4-aminopyridine.

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Cell Physiol 281:C290-C299.

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# TABLE 11 $K_V 2.2$ channels

Channel name  $K_V 2.2^{1-3}$ 

Description Voltage-gated potassium channel, delayed rectifier

Other names CDRK

Molecular information Human: 911 aa, NM\_004770, chr. 8q13.2, KCNB2, GeneID: 9312, PMID: 9612272<sup>15</sup>

Mouse: 758 aa, XM\_136482 (predicted), chr. 1

Rat: 802 aa, NM\_054000, chr. 5q11

Associated subunits Mouse K<sub>VB</sub>4 associates with K<sub>V</sub>2.2 and enhances expression level, K<sub>V</sub>8.1, K<sub>V</sub>9, KChaP<sup>4-7</sup>

Functional assays Voltage-clamp Current None determined

 $\begin{array}{lll} Conductance & 15 pS^8 \\ Ion \ selectivity & K^+\text{-selective} \\ Activation & Voltage \\ Inactivation & Noninactivating \end{array}$ 

Activators None Gating inhibitors None

Blockers Quinine (13.7  $\mu$ M), tetraethyammonium (2.6 mM), 4-aminopyridine (1.5 mM), phencyclidine ( $\mu$ M)<sup>8,9</sup>

Radioligands None

Channel distribution Brain [olfactory bulb (granule cell layer > olfactory tubercle) > cortex > hippocampus > cerebellum;

hypothalamus], ventricle, tongue, sympathetic neurons, gastrointestinal smooth muscle,

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mesenteric artery smooth muscle<sup>1-3,10-14</sup>

Physiological functions Maintaining membrane potential, modulating electrical excitability in neurons

Mutations and pathophysiology
Pharmacological significance
Not established
Not established

Comments The angiotensin II type 1 receptor mediates inhibition of  $K_v2.2$  in brainstem and hypothalamic

neurons<sup>12</sup>; mammalian *Shab*-related family

aa, amino acids; chr., chromosome.

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### TABLE 12 $K_{\rm v}3.1$ channels

 $K_V3.1$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names

Kv3.1, NGK2, KV4, KShIIIB, KS Molecular information

Mouse: 511aa, NM\_008421, chr. 7 Rat: 585aa, NM\_012856, chr. 1q22

Associated subunits Not established Functional assays Electrophysiology Current Delayed rectifier

 $27pS^{1,5}$ Conductance

Ion selectivity  $K^{+}(1) > Rb^{+}(0.76) > NH_{4}^{+}(0.12) = Cs^{+}(0.12) > Na^{+}(0.004)^{6}$ 

 $V_{\rm a} = 16 \text{ mV}; k_{\rm a} = 10 \text{ mV}; \tau_{\rm a} = 2 \text{ ms} (40 \text{ mV})^7$ Activation

 $\tau_{\rm h} = 630 \text{ ms} (40 \text{ mV})^1$ Inactivation

Activators None Gating inhibitors None

Blockers 4-Aminopyridine (29  $\mu$ M), capsaicin (158  $\mu$ M), resiniferatoxin (46  $\mu$ M), flecainide (108  $\mu$ M),

nifedipine (131 μM), diltiazem (97 μM), cromakalim (237 μM), tetraethyammonium (0.2 mM)<sup>8</sup>

Radioligands

Channel distribution Brain (cerebellum > globus pallidus, subthalamic nucleus, substantia nigra > reticular thalamic nuclei, cortical and hippocampal interneurons > inferior colliculi, cochlear and vestibular nuclei),

skeletal muscle, human Louckes B cells, germ cell, lung, testis, AtT20 cell line<sup>9–13,19,20</sup>

Important for the high-firing frequency of auditory<sup>8</sup> and fast-spiking GABAergic interneurons<sup>11,21</sup>; Physiological functions

regulation of action potential duration in presynaptic terminals 17,18 Kv3.1-/- mice exhibit impaired motor skills and reduced muscle contraction force<sup>13</sup>; Kv3.1/Kv3.3 Mutations and pathophysiology

double knockout mice display severe ataxia, myoclonus, and hypersensitivity to ethanol<sup>14</sup>

Not established

H-ras oncogene switches anterior pituitary-derived cells (AtT20) to a more neuron-like phenotype in parallel with the induction of expression of K<sub>V</sub>3.1<sup>12</sup>; mammalian Shaw-related family

aa, amino acids; chr., chromosome

Pharmacological significance

Comments

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TABLE 13  $K_{\rm V}3.2$  channels

 $K_{\rm V}3.2$ Channel name

Description Voltage-gated potassium channel, delayed rectifier RKShIIIA,1 Raw1,2 Kv3.2a,3 rKv3.2b and rKv3.2c4 Other names

Human: 613aa, NM\_139136 (transcript variant 1), chr. 12q14.1, KCNC2, GeneID: 3747, PMID: Molecular information

 $8111118^{21}$ 

Mouse: AC121610 (genomic), chr. 10

Rat: 613aa, NM\_139216 (transcript variant a), chr. 7q12-22

Associated subunits

Functional assays Electrophysiology Current Delayed rectifier Conductance  $16-20pS^{16}$ 

Ion selectivity  $K^+$ 

Activation  $V_{\rm a} = 13~{\rm mV}; \, k_{\rm a} = 7 - 7.5~{\rm mV^1}; \, t_{\rm on} = 10 - 90\% \; (40~{\rm mV}) \; 4~{\rm ms}; \; \tau_{\rm off} \; 2.9 \; {\rm ms} \; (-60~{\rm mV})^{16}$ 

Inactivation Activators None Gating inhibitors None

Tetraethyammonium (0.1 mM),<sup>6</sup> 4-aminopyridine (0.1 mM),<sup>6</sup> 8-bromo-cGMP,<sup>7</sup> 3-isobutyl-1-Blockers

methylxanthine,<sup>6</sup> D-NONOate,<sup>7</sup> verapamil (11 μM),<sup>8</sup> ShK<sup>19</sup>

Radioligands

Channel distribution Brain (fast-spiking GABAergic interneurons of the neocortex, hippocampus, and caudate; terminal

fields of thalamocortical projections), 9-12 islets, 13 mesenteric artery, Schwann cells 14

Physiological functions Probably in heteromeric complexes with K<sub>v</sub>3.1; important for the high-frequency firing of fast

spiking GABAergic interneurons<sup>17</sup> and GABA release via regulation of action potential duration in presynaptic terminals<sup>18</sup>; modulated by protein kinase A in vitro and in vivo<sup>10,20</sup>

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See "Comments' Mutations and pathophysiology

Pharmacological significance Not established

Comments Fast deactivation; knockout mice show specific alterations in their cortical electroencephalographic patterns and an increased susceptibility to epileptic seizures consistent with an impairment of a

cortical inhibitory mechanism<sup>15</sup>; mammalian Shaw-related family

aa, amino acids; chr., chromosome; D-NONOate, 1,1-diethyl-2-hydroxy-2-nitrosohydrazine; ShK,  $Stychodactyla\ helianthus\ toxin$ . 1. McCormack T, Vega-Saenz de Miera EC, and Rudy B (1991) Molecular cloning of a member of a third class of Shaker-family K $^+$  channel genes in mammals.  $Proc\ Natl$ Acad Sci USA 87:5227-5231.

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distinct membrane targeting signals. J Membr Biol 159:149-159.

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HARMACOLOGICAL REVIEW

TABLE 14
$K_{\rm v}3.3$ channels

 $K_V 3.3^{1-4}$ Channel name

Description Voltage-gated A-type potassium channel<sup>2</sup> Other names hKv3.3, mKv3.3,1 RKShIIID,3 Kv3.3b4

Human: 757aa, NM\_004977, chr. 19q13.3-4, 1-3 KCNC3, GeneID: 3748, PMID: 17403291 Molecular information

> Mouse: 679aa, NM\_008422, chr. 7 Rat: 770aa, NM\_053997, chr. 1q22

Associated subunits None

Functional assays Electrophysiology

Current A-type

Conductance Not established

Ion selectivity

 $V_{\rm a} = 7 \text{ mV}; k_{\rm a} = 6 \text{ mV}^2$ Activation  $\tau_h \sim \, 200 \ ms \ (40 \ mV)^2$ Inactivation

Activators None Gating inhibitors None

Tetraethyammonium (0.14 mM),<sup>2</sup> 4-aminopyridine (1.2 mM)<sup>2</sup>; blocked by hypoxia<sup>5</sup> Blockers

Radioligands

Brain, Purkinje cells, central nervous system motoneurons; auditory brainstem<sup>12</sup>; electrosensory, Channel distribution

cerebellar neurons, central auditory nuclei<sup>6-8</sup>; mesenteric artery<sup>9</sup>; lens and corneal epithelium<sup>10</sup>

Physiological functions Not established Mutations and pathophysiology See "Comments' Pharmacological significance Not established

Comments Alcohol hypersensitivity, ataxia, increased locomotion and myoclonus occur in mice lacking  $K_V3.3$ 

and K<sub>v</sub>3.1<sup>11</sup>; mammalian Shaw-related family

aa, amino acids; chr., chromosome.

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# TABLE 15 $K_{\nu}3.4$ channels

Channel name K<sub>V</sub>3.4

Description Voltage-gated potassium channel, A-type, fast-inactivating

Other names Raw3, HKShIIIC, MKv3.43

Molecular information Human: 635 aa, NM\_004978 (transcript variant 1), chr. 1p21<sup>1,2</sup>, KCNC4, GeneID: 3749, PMID:

 $1920536^2$ 

Mouse: 628 aa, NM\_145922, chr. 3

Rat:

Associated subunits MiRP2 forms potassium channels in skeletal muscle with  $K_v3.4^4$ 

Functional assays Electrophysiology

 $\begin{array}{ll} Current & A-type \\ Conductance & 14pS^{1,5} \\ Ion \ selectivity & K^+ \end{array}$ 

Activation  $V_a = 3.4 \text{ mV}^5, +14 \text{ mV}^1; k_a = 8.4 \text{ mV}^5$ 

Inactivation N-type inactivation,  $V_h = 53$  mV;  $k_h = 7.4$  mV;  $\tau_h = 15.9$  ms (50 mV)<sup>1,2,5</sup>

Activators None Gating inhibitors None

Blockers BDS-I (47 nM),<sup>6</sup> tetraethyammonium (0.3 mM)<sup>1,5</sup>; the specificity of BDS-I for K<sub>v</sub>3.4 has been

 $questioned^{12}$ 

Radioligands None

Channel distribution Parathyroid, prostate, brain (brainstem, hippocampal granule cells), skeletal muscle, 48.9

pancreatic acinar cells<sup>10,11</sup>

Physiological functions Together with MirP2 forms low-voltage-ctivating potassium channels that regulate skeletal muscle

resting potential4

Mutations and pathophysiology Mutations of MiRP2, which associates with  $K_v3.4$  in skeletal muscle, are associated with periodic

utations of Mi paralysis<sup>4</sup>

Pharmacological significance Not established

Comments Mammalian Shaw-related family

aa, amino acids; chr., chromosome.

1. Schroter KH, Ruppersberg J, Wunder F, Rettig J, Stocker M, and Pongs O (1991) Cloning and functional expression of a TEA-sensitive A-type potassium channel from rat brain. FEBS Lett 278:211–216.

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TABLE 16
$K_{\rm v}$ 4.1 channels

Channel name  $K_{V}4.1$ 

Description Voltage-gated potassium channel, A-type potassium current

Other names mShal1

Human: 647aa, NM\_004979, chr. Xp11.23,2 KCND1 (see 'Comments'), GeneID: 3750, PMID: Molecular information

 $10729221^{12}$ 

Mouse: 651aa, NM\_008423, chr. X

Rat: 650aa, XM\_217601 (predicted), chr. Xq13

Associated subunits KChIP1 increases  $K_V4.1$  current densities, accelerates inactivation time course and recovery from

inactivation, and shifts steady-state inactivation to more depolarized potentials<sup>3,4</sup>

Functional assays Patch-clamp, two-electrode voltage-clamp

Current Somatodendritic depolarization-activated potassium currents in rat neostriatal cholinergic

interneurons are predominantly of the A-type and attributable to coexpression of K<sub>V</sub>4.2 and K<sub>V</sub>4.1

subunits<sup>5</sup>; subthreshold transient A currents in rat brain<sup>6</sup> ~6pS (main unitary conductance under physiological conditions)<sup>4,7</sup>

Ion selectivity  $P_{Na}/P_{K} < 0.01$ 

Activation

Na R Voltage,  $V_a=-47.9$  mV;  $k_a=24.2$  mV (assuming a fourth-order Boltzmann function)<sup>7</sup>  $V_h=-69$  mV;  $k_h=4.8$  mV;  $\tau_{h1}=22$  ms (20 mV);  $\tau_{h2}=86$  ms (20 mV);  $\tau_{h3}=368$  ms (20 mV)<sup>7</sup> (see "Comments") Inactivation

None Activators Gating inhibitors None

4-Aminopyridine (9 mM)<sup>1,7</sup>, tetraethyammonium (>10 mM)<sup>1</sup> Blockers

Radioligands

Conductance

Channel distribution Fetal, infant, and adult brain; colon, heart, lung, stomach, testis, liver, kidney, thyroid gland,

pancreas, pulmonary artery8-10

Physiological functions Not established Mutations and pathophysiology Not established Pharmacological significance Not established

Comments The  $K_n4.1$  (KCND1) gene is encoded by at least 6 exons<sup>2</sup>—the first exon encodes the protein from the N terminus through S5 into the P-region, whereas the remainder of the protein is encoded by

exons 2-6; kinetic properties depend on the expression system, recording configuration, and the

presence of auxiliary subunits (KChIPs)<sup>4,11</sup>; mammalian Shal-related family

aa, amino acids; chr., chromosome.

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potassium channels mediating A-type currents  $I_{TO}$  and  $I_{SA}$ . Genomics **64**:144–154.

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TABLE 17  $K_{\rm V}$ 4.2 channels

 $K_{\rm V}4.2$ Channel name

Description Voltage-gated potassium channel, A-type potassium current

Other names Shal1, RK5<sup>1-3</sup>

Human: 630aa, NM\_012281, chr. 7q31, KCND2 (see "Comments"), GeneID: 3751, PMID: 10551270<sup>24</sup> Molecular information

> Mouse: 630aa, NM\_019697, chr. 6 Rat: 490aa, NM\_031730, chr. 4q22

Associated subunits

Coexpression of KChIP1 results in increased current densities, slowed onset of inactivation, and accelerated recovery from inactivation<sup>4</sup>; KChIP4/CALP interacts with K<sub>V</sub>4.2 and presentilin 2<sup>5</sup>; frequenin, a calcium-binding protein, enhances K<sub>v</sub>4.2 current amplitudes, slows inactivation time course and accelerates recovery from inactivation<sup>6</sup>; PSD95, a PDZ domain protein, associates with  $K_V4.2$  and is involved in trafficking of the channel<sup>7</sup>; a number of proteins have been shown to interact and modify K<sub>v</sub>4 proteins, including KChIPs, DPPX, DPP10, frequenin, PSD95, and filamin—most of these studies have used K<sub>v</sub>4.2 and sometimes K<sub>v</sub>4.3 proteins, but it is likely that these interactions also occur with Kv4.1; the physiological role of these proteins in native channels remains to be studied in most cases

Functional assays Patch-clamp, two-electrode voltage-clamp

Current  $I_{to}$  current in the heart is a heteromultimer of  $K_V4.2$  and  $K_V4.3$  subunits and KChIP28;  $I_{SA}$  current

in somatic recordings from neurons9

Conductance Not established Ion selectivity  $P_{\rm Na}\!/P_{\rm K} < 0.01$ 

Midpoint of activation =  $1 \text{ mV}^2$ Activation

Inactivation Rapid inactivation with time constants of 15 and 60 ms<sup>2</sup>

Activators None Gating inhibitors None

4-Aminopyridine (5 mM), 1,10 heteropodatoxins, 11 PaTX1,2 (2-70 nM), arachidonic acid (2 \(\mu M\))^{12} Blockers

Radioligands None

Channel distribution Brain [cerebellum (granular cells) > hippocampus, thalamus, medial habenular nucleus > cerebral cortex; basal ganglia and forebrain<sup>13</sup>; concentrated in dendrites and soma<sup>14</sup>], cochlear nucleus, <sup>15</sup> atrium, ventricle 1-3,16; in situ hybridization has shown that many neuronal populations preferentially express  $K_V4.2$  or  $K_V4.3^{23}$ —for example, CA1 hippocampal neurons express  $K_V4.2$ but not K<sub>V</sub>4.3—on the other hand, Purkinje cells and cortical interneurons express K<sub>V</sub>4.3 preferentially; in cerebellar granule cells, there is a reciprocal anterior-posterior gradient of

expression

Physiological functions Repolarization of the cardiac action potential (notch phase), dampening back-propagating action potentials in CA1 hippocampal neurons

KChIP2-/- mice lack the  $I_{to}$  current and are susceptible to ventricular tachycardia<sup>17</sup>; seizure Mutations and pathophysiology

activity reduces K<sub>V</sub>4.2 expression in the dentate granule cells of the hippocampus<sup>18</sup>

Not established

The K.4.2 (KCND2) gene, like KCND1 and KCND3, contains six exons—however, the introns are significantly longer<sup>19</sup>; kinetic properties depend on the expression system, recording configuration, and the presence of auxiliary subunits (KChIPs) $^{20,21}$ ;  $K_v4.2$  currents expressed in *Xenopus* oocytes are suppressed in response to protein kinase C activation<sup>22</sup>; mammalian Shal-related family

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aa, amino acids; chr., chromosome.

Pharmacological significance

Comments

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### TABLE 18 $K_{\rm V}4.3$ channels

 $K_{\rm V}4.3^{1-6}$ Channel name

Description Voltage-gated potassium channel, A-type potassium current

Other names

Molecular information Human: 655aa, NM\_004980 (transcript variant 1), chr. 1p13.3, KCND3 (see "Comments"), GeneID:

3752, PMID: 8734615<sup>2</sup>

Mouse: 655aa, NM\_019931, chr. 3 Rat: 636aa, NM\_031739, chr. 2q34

KChIP1 increases K<sub>v</sub>4.3 current densities, accelerates inactivation time course and recovery from Associated subunits

inactivation, and shifts steady-state inactivation to more depolarized potentials; KChIP4a abolishes fast inactivation<sup>7</sup>; expression of  $K_{VB}2$  in brain increases current density and protein expression<sup>8</sup>; KChAP acts as a chaperone for K<sub>V</sub>4.3<sup>9</sup>; K<sub>V</sub>4.3 may associate preferentially with DPP10 in native neurons that predominantly express this subunit<sup>20</sup>

Functional assays Patch-clamp, two-electrode voltage-clamp

 $I_{to}$  current in the heart is a heteromultimer of  $K_V4.2$  and  $K_V4.3$  subunits and KChIP210 Current

~5pS (main unitary conductance under physiological conditions)<sup>7</sup>; association with DPPX increases Conductance

single channel conductance<sup>21</sup>

Ion selectivity  $P_{Na}/P_{K} < 0.01$ 

Threshold for activation -30 mV, time course for activation 1.71 ms at 60 mV<sup>11</sup> Activation

Inactivation Time course for inactivation fit by a biexponential function;  $\tau_{h1} = 27$  ms at 60 mV,  $\tau_{h2} = 142$  ms at

60 mV<sup>11</sup> (see "Comments")

Activators None Gating inhibitors None

4-Aminopyridine, bupivacaine (31 μM), <sup>11</sup> PaTX1,2, (2–70 nM), nicotine (40 nM)<sup>12</sup> Blockers

Radioligands

Heart, brain, smooth  $muscle^{1-6,13,14}$ Channel distribution

Physiological functions Repolarization of the cardiac action potential (notch phase)

K<sub>v</sub>4.3 mRNA levels are decreased in patients with paroxysmal atrial fibrillation<sup>15</sup> Mutations and pathophysiology

Pharmacological significance

The  $K_04.3$  (KCND3) gene contains six exons analogous to those found in KCND1 and KCND2 and an additional exon L between exons 4 and 5—relative to KCND1, the introns are significantly longer; kinetic properties depend on the expression system, recording configuration, and the presence of auxiliary subunits (KChIPs)<sup>16–18</sup>; K<sub>v</sub>4.3 currents expressed in Xenopus oocytes are suppressed in response to protein kinase C activation<sup>19</sup>; mammalian Shal-related family

aa, amino acids; chr., chromosome

Comments

1. Serodio P, Kentros C, and Rudy B (1994) Identification of molecular components of A-type channels activating at subthreshold potentials. J Neurophysiol **72:**1516-1529.

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- 21. Rocha CA, Nadal M, Rudy B, and Covarrubias M. (2004) Inactivation gating of Kv4 K<sup>+</sup> channels interacting with the dipeptidyl-aminopeptidase-like protein (DPPX), in *Proceedings of the 48th Annual Meeting of the Biophysical Society*; 2004 14–18 Feb; Baltimore, Md. Presentation 2780-Pos.

### TABLE 19 $K_{V}5.1$ channels

Channel name  $K_V 5.1^{1-4}$ 

Description Modifier of the  $K_V2$  family of channels

Other names KH1, IK8

Molecular information Human: 494aa, NM\_002236, chr. 2p25, KCNF1, GeneID: 3754, PMID: 9434767<sup>5</sup>

Mouse: 493aa, NM\_201531, chr. 12 Rat: 505aa, XM\_216678 (predicted), chr. 6

Associated subunits Associates with  $K_v2.1$  and  $K_v2.2$ 

Functional assays Voltage-clamp

Current None

Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Brain, heart, skeletal muscle, liver, kidney pancreas, <sup>1,2,6</sup> cardiac myocytes<sup>7</sup>

Physiological functions Modifies the gating properties of  $K_V 2.1$  and  $K_V 2.2$  channels

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments  $K_V$ 5.1 has no function on its own, but it has important modulatory actions on  $K_V$ 2 channels

aa, amino acids; chr., chromosome.

- 1. Drewe JA, Verma S, Frech G, and Joho RH (1992) Distinct spatial and temporal expression patterns of K<sup>+</sup> channel mRNAs from different subfamilies. *J Neurosci* 12:538–548.
- 2. Verma-Kurvari S, Border B, and Joho RH (1997) Regional and cellular expression patterns of four K<sup>+</sup> channel mRNAs in the adult rat brain. Brain Res Mol Brain Res 6:54–62.
- 3. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian Shab K<sup>+</sup> channels. J Biol Chem 272:24371–24379.

  4. Kramer JW, Post MA, Brown AM, and Kirsch GE (1998) Modulation of potassium channel gating by coexpression of Kv2.1 with regulatory Kv5.1 or Kv6.1 α-subunits. Am J Physiol 274:C1501–C1510.
- 5. Su K, Kyaw H, Fan P, Zeng Z, Shell BK, Carter KC, and Li Y (1997) Isolation, characterization, and mapping of two human potassium channels. Biochem Biophys Res Commun 241:675–681.
  - 6. UniGeneCluster Hs0.23735; OMIM no. 603787.
- 7. Brahmajothi MV, Morales MJ, Liu S, Rasmusson RL, Campbell DL, and Strauss HC (1996) In situ hybridization reveals extensive diversity of K<sup>+</sup> channel mRNA in isolated ferret cardiac myocytes. Circ Res 78:1083–1089.



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# TABLE 20 $K_{\rm V}$ 6.1 channels

Channel name  $K_V 6.1^{1-6}$ 

Description Modifier/silencer of  $K_V2$  family channels

Other names KH2, K13

Molecular information Human: 513aa, NM\_002237, chr. 20q13, 6.7 KCNG1, GeneID: 3755, PMID: 94347676

Mouse: 534aa, XM\_141545 (predicted), chr. 2 Rat: 514aa, XM\_215951 (predicted), chr. 3 Associates with  $K_{\rm V}2$  family channels

Functional assays Electrophysiology

Current None

Associated subunits

Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Skeletal muscle, brain, uterus, ovary, kidney, pancreas, placenta, bone, germ cell, prostate, skin,

testis, <sup>6,7</sup> cardiac myocytes (sinoatrial node)<sup>8</sup>

Physiological functions  $K_v6.1$  subunits when expressed alone are unable to elicit any current—however,  $K_v6.1$  can

suppress  $K_V2.1$  current (less effectively than  $K_V5.1$ ), and to a lesser extent it can suppress  $K_V2.2$ ; the  $K_V2.1$  currents are strongly modified by  $K_V6.1$ , which increases the time constant of activation

and slows down inactivation

Mutations and pathophysiology Pharmacological significance

Not established Not established

Comments  $K_V6.1$  has no function on its own, but it has important modulatory actions on  $K_V2$  channels

aa, amino acids; chr., chromosome.

- 1. Drewe JA, Verma S, Frech G, and Joho RH (1992) Distinct spatial and temporal expression patterns of K<sup>+</sup> channel mRNAs from different subfamilies. J Neurosci 12:538–548.
- 2. Post MA, Kirsch GE, and Brown AM (1996) Kv2.1 and electrically silent Kv6.1 potassium channel subunits combine and express a novel current. FEBS Lett 399:177–182.
- 3. Verma-Kurvari S, Border B, and Joho RH (1997) Regional and cellular expression patterns of four K<sup>+</sup> channel mRNAs in the adult rat brain. *Brain Res Mol Brain Res* 46:54–62.
- 4. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunsk, M (1997) New modulatory α subunits for mammalian Shab K<sup>+</sup> channels. J Biol Chem 272:24371–24379. 5. Kramer JW, Post MA, Brown AM, and Kirsch GE (1998) Modulation of potassium channel gating by coexpression of Kv2.1 with regulatory Kv5.1 or Kv6.1 α-subunits.
- Am J Physiol 274:C1501–C1510.
  6. Su K, Kyaw H, Fan P, Zeng Z, Shell BK, Carter KC, and Li Y (1997) Isolation, characterization, and mapping of two human potassium channels. Biochem Biophys Res Commun 241:675–681.
  - 7. UniGene Cluster Hs0.118695; OMIM no. \*603788.
- 8. Brahmajothi MV, Morales MJ, Liu S, Rasmusson RL, Campbell DL, and Strauss HC (1996) In situ hybridization reveals extensive diversity of K<sup>+</sup> channel mRNA in isolated ferret cardiac myocytes. Circ Res 78:1083–1089.

# TABLE 21 $K_V 6.2$ channels

 $\begin{array}{ll} \text{Channel name} & \quad K_{V}6.2^{1} \\ \text{Description} & \quad \text{Modifier/silencer} \end{array}$ 

Other names None

Molecular information Human: 466aa, NM\_012283, chr. 18q22-18q23, <sup>1</sup> KCNG2, GeneID: 26251, PMID: 10551266<sup>1</sup>

Mouse: AC145610 (genomic), chr. 18 Rat: 436aa, XM\_225718 (predicted), chr. 18

Associated subunits Coassembles with  $K_V2$  family channels via the N termini<sup>1</sup>

Functional assays Electrophysiology

Current None

Conductance Not functional on its own
Ion Selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Myocardium, fetal brain, germinal center B cells<sup>1,2</sup>

 $K_{v}6.3^{1}$ 

Modifier/silencer

Not established

Physiological functions Modifier/silencer, coassembles with K<sub>V</sub>2.1, producing K<sup>+</sup> channels with unique properties

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments  $K_V 6.2$  has no function on its own, but it has important modulatory actions on  $K_V 2$  channels

aa, amino acids; chr., chromosome.

Channel name

Description

### TABLE 22 $K_{v}6.3$ channels

Other names	$\mathrm{K_{v}}10.1$
Molecular information	Human: 436aa, NM_133329, chr. 2p21, KCNG3, GeneID: 170850, PMID: 11852086 <sup>1</sup>
	Mouse: 433aa, NM_153512, chr. 17
	Rat: 345aa, NM_133426, chr. 6q12
Associated subunits	Coassembles with ${ m K_V}2.1^1$
Functional assays	Electrophysiology
Current	None
Conductance	Not functional on its own
Ion selectivity	Not functional on its own
Activation	Not functional on its own
Inactivation	Not functional on its own
Activators	None
Gating inhibitors	None
Blockers	None
Radioligands	None
Channel distribution	Whole brain (hippocampus, caudate nucleus, frontal lobe, hypothalamus, substantia nigra), spinal cord, pituitary, testis, small intestine, thymus, adrenal gland <sup>1</sup>
Physiological functions	Modifier/silencer, coassembles with ${ m K_V}2.1$
Mutations and pathophysiology	Not established

aa, amino acids; chr., chromosome.

Pharmacological significance

Comments

 $K_v6.3$  has no function on its own, but it has important modulatory actions on  $K_v2$  channels



<sup>1.</sup> Zhu XR, Netzer R, Bohlke K, Liu Q, and Pongs O (1999). Structural and functional characterization of Kv6.2: a new γ-subunit of voltage-gated potassium channel. Receptors Channels 6:337–350.

<sup>2.</sup> UniGene Cluster Hs0.247905; OMIM no. 605696.

<sup>1.</sup> Sano Y, Mochizuki S, Miyake A, Kitada C, Inamura K, Yokoi H, Nozawa K, Matsushime H, and Furuichi K (2002) Molecular cloning and characterization of Kv6.3, a novel modulatory subunit for voltage-gated K<sup>+</sup> channel Kv2.1. FEBS Lett **512**:230–234.

# Spet

# TABLE 23 $K_{v}6.4$ channels

Channel name	$ m K_V 6.4^1$
Channel name	$\mathrm{K_{V}6.4^{1}}$

Description Modifier/silencer

Other names None

Molecular information Human: 519aa, NM\_172347 (transcript variant 1), chr. 16q24.1, KCNG4, GeneID: 93107, PMID:

 $12060745^{1}$ 

Mouse: 506aa, NM\_025734, chr. 8,

Rat: 506aa, XM\_226524 (predicted), chr. 19

Associated subunits  $Coassembles with K_V 2.1^1$ 

Functional assays Electrophysiology

Current Not functional on its own
Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Brain, liver, small intestine, colon<sup>1</sup>

Physiological functions Regulation of membrane potential and action potential frequency by modulation of delayed rectifier

potassium currents; modulates the activity of K<sub>V</sub>2.1 channels by causing marked changes in

activation threshold and kinetics, C-type inactivation, and deactivation<sup>1</sup>

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments  $K_V6.4$  has no function on its own, but it has important modulatory actions on  $K_V2$  channels

aa, amino acids; chr., chromosome.

<sup>1.</sup> Ottschytsch N, Raes A, Van Hoorick D, and Snyders DJ (2002) Obligatory heterotetramerization of three previously uncharacterized Kv channel-subunits identified in the human genome. Proc Natl Acad Sci USA 99:7986–7991.

TABLE 24  $K_{\rm v}7.1\ channels$ 

Channel name K<sub>V</sub>7.1

Description Voltage-gated potassium channel, delayed rectifier

Other names KVLQT1, slow delayed rectifier

Molecular information Human: 676aa, NM\_000218 (transcript variant 1), chr. 11p15.5, KCNQ1, GeneID: 3784, PMID:

 $8528244^{1}$ 

Mouse: 668aa, NM\_008434, chr. 7 Rat: 669aa, NM\_032073, chr. 1q41

Associated subunits KCNE1 (minK/IsK), KCNE3 [minK-related peptide 2 (MiRP2)]

Functional assays Voltage-clamp

 $\begin{array}{ll} {\rm Current} & {\rm IK_s~(with~KCNE1)}, ^{2.3}~{\rm IK_{cAMP}~(with~KCNE3)}^{16} \\ {\rm Conductance} & {\rm 1.8pS~(KCNQ1~alone),~5pS~(with~KCNE1)} \end{array}$ 

Ion selectivity K<sup>+</sup>

Activation KCNQ1 alone:  $V_a = 12$  mV,  $\tau_a = 30$ , and 800 ms at +40 mV

KCNQ1 + KCNE1:  $V_{\rm a}$  = +8 mV,  $\tau_{\rm a}$  = 0.7, 1.5, and 8 s at +40 mV

Inactivation KCNQ1 alone:  $V_{\rm h}$  = +18 mV,  $\tau_{\rm h}$  = 130 ms at 20 mV

Activators R-L3 (= L364373, 1  $\mu$ M for KCNQ1 alone; R-L3 does not activate the KCNQ1/KCNE1 complex; the

S enantiomer blocks KCNQ1)4; mefenamic acid, niflumic acid, and 4,4'-diisothiocyanostilbene-2,2'-

disulfonic acid (10–100  $\mu\mathrm{M})^{5,6}$ 

Gating inhibitors None

Blockers Chromanol 293B (1  $\mu$ M), L735821 (80 nM), mefloquine (0.88  $\mu$ M), azimilide (3  $\mu$ M), similide (3  $\mu$ M), hMR-

1556 (120 nM), XE991 (0.78 μM KCNQ1 alone; 11.1 μM KCNQ1/KCNE1), 11 linopirdine (8.9 μM

KCNQ1 alone)

Radioligands None

Channel distribution Heart, kidney, rectum, ear, germ, pancreas, lung, cochlea, placenta

Physiological functions Repolarization of cardiac action potentials (KCNQ1 and minK/ISK/KCNE1 coassemble to form the cardiac IK<sub>s</sub> channel); potassium recycling at basolateral membrane of intestinal crypt cells (with

KCNE3) and inner ear

Mutations and pathophysiology Loss of function mutations in the KCNQ1 gene can cause either RWS (autosomal dominant) or

JLNS (autosomal recessive); RWS is characterized by congenital long QT syndrome and electrocardiographically distinguished by a prolonged QT interval and polymorphic ventricular arrhythmias (torsade de pointes), which may result in recurrent syncopes, seizure, or sudden death; JLNS patients have deafness, congenital and functional heart disease, a prolonged QT interval on an electrocardiogram, and sudden death cardioauditory syndrome; KCNQI is disrupted by chromosomal rearrangements in patients with Beckwith-Wiedemann syndrome, <sup>13</sup> as well as by a balanced chromosomal translocation in an embryonal rhabdoid tumor; gain-of-

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function mutations in KCNQ1 cause atrial fibrillation and short QT syndrome

Pharmacological significance Blockers developed as class III antiarrhythmic agents to target ventricular arrhythmias 14,15;

activators could be useful for the treatment of some long QT syndromes<sup>6</sup>

aa, amino acids; chr., chromosome; RWS, Romano-Ward syndrome; JLNS, Jervell and Lange-Nielsen syndrome; L735821, 3-(2,4-dichlorophenyl)-N-(6-methyl-5-oxo-2-phenyl-3,6-diazabicyclo[5.4.0]undeca-2,7,9,11-tetraen-4-yl)-prop-2-enamide; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; HMR-1556, N-(6-cyano-3-hydroxy-2,2-dimethyl-chroman-4-yl)N-methyl-ethansesulfonamide.

1. Wang Q, Curran ME, Splawski I, Burn TC, Millholland JM, Van Raay TJ, Shen J, Timothy KW, Vincent GM, de Jager T, et al. (1996) Positional cloning of a novel potassium channel gene: KVLQT1 mutations cause cardiac arrhythmias. Nat Genet 12:17–23.

2. Sanguinetti MC, Curran ME, Zou A, Shen J, Spector PS, Atkinson DL, and Keating MT (1996) Coassembly of K<sub>V</sub> LQT1 and minK (IsK) proteins to form cardiac I<sub>Ks</sub> potassium channel. Nature (Lond) 384:80-83.

3. Barhanin J, Lesage F, Guillemare E, Fink M, Lazdunski M, and Romey G (1996) K<sub>V</sub> LQT1and lsK (minK) proteins associate to form the I<sub>Ks</sub> cardiac potassium current. Nature (Lond) 384:78-80.

4. Salata JJ, Jurkiewicz NK, Wang J, Evans BE, Orme HT, and Sanguinetti MC (1998) A novel benzodiazepine that activates cardiac slow delayed rectifier K<sup>+</sup> currents. Mol Pharmacol 54:220–230.

5. Busch AE, Herzer T, Wagner CA, Schmidt F, Raber G, Waldegger S, and Lang F (1994) Positive regulation by chloride channel blockers of IsK channels expressed in Xenopus oocytes. Mol Pharmacol 46:750–753.

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8. Tinel N, Lauritzen I, Chouabe C, Lazdunski M, and Borsotto M (1998) The KCNQ2 potassium channel: splice variants, functional and developemental expression: brain localization and comparison with KCNQ3. FEBS Lett 438:171–176.

9. Kang J, Chen XL, Wang L, and Rampe D (2001) Interactions of the antimalarial drug mefloquine with the human cardiac potassium channels KvLQT1/minK and HERG. J Pharmacol Exp Ther 299:290–296.

- 10. Busch AE, Busch GL, Ford E, Suessbrich H, Lang HJ, Greger R, Kunzelmann K, Attali B, and Stuhmer W (1997) The role of the IsK protein in the specific pharmacological properties of the IKs channel complex. Br J Pharmacol 122:187–189.
- 11. Wang HS, Brown BS, McKinnon D, and Cohen, IS (2000) Molecular basis for differential sensitivity of KCNQ and I<sub>Ks</sub> channels to the cognitive enhancer XE991. *Mol Pharmacol* 57:1218–1223.
  - $12. \ Keating\ MT\ and\ Sanguinetti\ MC\ (2001)\ Molecular\ and\ cellular\ mechanisms\ of\ cardiac\ arrhythmias.\ \textit{Cell}\ 104:569-580.$
- 13. Lee MP, Hu RJ, Johnson LA, and Feinberg AP (1997) Human KVLQT1 gene shows tissue-specific imprinting and encompasses Beckwith-Wiedemann syndromechromosomal rearrangements. Nat Genetics 15:181–185.
- 14. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent develo pMents in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. J Med Chem 44:1627–1653.
- 15. Shieh CC, Coghlan M, Sullivan JP, and Gopalakrishnan M (2000) Potassium channels: molecular defects, diseases, and therapeutic opportunities. *Pharmacol Rev* 52:557–594.
- 16. Schroeder BC, Waldegger S, Fehr S, Bleich M, Warth R, Greger R, and Jentsch TJ (2000) A constitutively open potassium channel formed by KCNQ1 and KCNE3. Nature (Lond) 403:196–199.

### TABLE 25 $K_{\rm V}7.2$ channels

 $K_V7.2$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names

Molecular information Human: 872aa, NM\_172107 (transcript variant 1), chr. 20q13.3, KCNQ2, GeneID: 3785, PMID:

 $9836639^{1}$ 

Mouse: 870aa, NM\_010611 (transcript variant 1), chr. 2

Rat: 852aa, NM\_133322, chr. 3q43

Associated subunits KCNQ3, KCNE2 Functional assays Voltage-clamp Current M current Conductance  $5.8 pS^{13}$ Ion selectivity

Activation  $V_{\rm a}$  = 26 mV,  $\tau_{\rm a}$  = 157 ms at +30 mV Inactivation  $V_{\rm h}$  = 18 mV,  $\tau_{\rm h}$  = 130 ms at 20 mV Activators Retigabine (10  $\mu$ M), BMS204352 (1  $\mu$ M)<sup>3</sup>

Gating inhibitors

Blockers Tetraethyammonium (KCNQ2 alone: 0.16 mM; KCNQ2/KCNQ3: 0.5 mM), XE991 (0.7 μM), 1,4

linopiridine (4.8  $\mu$ M), <sup>1,3</sup> L735821 (1.5  $\mu$ M)<sup>5</sup>

Radioligands None

Channel distribution Infant brain, adult brain, fetal brain, sympathetic ganglia, lung, testis, fetal heart, adult heart,

breast, eye, germ cell, placenta, small intestine, neuroblastoma<sup>10</sup>

Physiological functions Determines subthreshold excitability of neurons; KCNQ2 and KCNQ3 coassemble to form the M current in the brain1 (see "Comments"); KCNQ2 and KCNQ3 proteins are colocalized in a

somatodendritic pattern on pyramidal and polymorphic neurons in the human cortex and hippocampus<sup>11</sup>; KCNQ2 is also expressed in the absence of KCNQ3 in some presynaptic terminals<sup>11</sup>

Benign familial neonatal convulsions (EBN1) with myokymia<sup>6,7</sup>; in KCNQ2 knockout mice, Mutations and pathophysiology

homozygotes (KCNQ2-/-) die within a few hours after birth owing to pulmonary atelectasis that is not due to the status of epileptic seizures, although their development is morphologically normal; heterozygous mice have decreased expression of KCNQ2 and show hypersensitivity to

pentylenetetrazole, an inducer of seizure<sup>12</sup> Pharmacological significance

Retigabine is an anticonvulsant<sup>2</sup> (the M current is a new target for antiepileptic therapy<sup>8,9</sup>);

blockers enhance learning and memory in animal models<sup>9</sup>

The M current is a slowly activating and deactivating potassium conductance that plays a critical Comments

role in determining the subthreshold excitability of neurons as well as the responsiveness to synaptic inputs; the M current was first described in peripheral sympathetic neurons, and differential expression of this conductance produces subtypes of sympathetic neurons with distinct firing patterns; the M current is also expressed in many neurons in the central nervous system

aa, amino acids; chr., chromosome; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1H-indol-2-one; XE991, 10,10-bis(pyridin-4-ylmethyl)anthra $cen-9-one;\ L735821,\ 3-(2,4-dichlor ophenyl)-N-(6-methyl-5-oxo-2-phenyl-3,6-diazabicyclo [5.4.0] undeca-2,7,9,11-tetraen-4-yl)-prop-2-enamide.$ 

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3. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, Olesen SP (2001) KCNQ4 channel activation by BMS-204352 and retigabine. Neuropharmacology

4. Robbins J (2001) KCNQ potassium channels: physiology, pathophysiology, and pharmacology. Pharmacol Ther 90:1–19.

5. Tinel N, Lauritzen I, Chouabe C, Lazdunski M, and Borsotto M (1998) The KCNQ2 potassium channel: splice variants, functional and develo pMental expression: brain localization and comparison with KCNQ3, FEBS Lett. 438:171-176.

6. Charlier C, Singh NA, Ryan SG, Lewis TB, Reus BE, Leach R, and Leppert M. (1998) A pore mutation in a novel KQT-like potassium channel gene in an idiopathic epilepsy family. Nat Genet 18:53-55.

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9. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent develo pMents in the biology and medicinal chemistry of potassium channel modulators: update from a decade of progress. J Med Chem 44:1627–1653.

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potassium channel subunits that are mutated in epilepsy. Proc Natl Acad Sci USA 97:4914-4919 12. Watanabe H, Nagata E, Kosakai A, Nakamura M, Yokoyama M, Tanaka K, and Sasai H (2000) Disruption of the epilepsy KCNQ2 gene results in neural hyperexcitability. J Neurochem 75:28-33.

13. Selyanko AA, Hadley JK, Wood IC, Abogadie FC, Delmas P, Buckley NJ, London B, and Brown DA (2001) Properties of single M-type KCNQ2/KCNQ3 potassium channels expressed in mammalian cells. J Physiol 534:15-24.



### TABLE 26 $K_{\rm V}7.3$ channels

 $K_V7.3$ Channel name

Voltage-gated potassium channel, delayed rectifier Description

Other names

Molecular information Human: 872aa NM\_004519, chr. 8q24, KCNQ3, GeneID: 3786, PMID: 9836639<sup>1</sup>

> Mouse: 873aa, NM\_152923, chr. 15 Rat: 873aa, NM\_031597, chr. 7q33

Associated subunits KCNQ2, KCNQ5 Functional assays Voltage-clamp M current<sup>1</sup> Current Conductance 7.3pSIon selectivity  $K^{\perp}$ 

 $V_{\rm a}$  = 39 mV,  $\tau_{\rm a}$  = 60 ms at +30 mV Activation

Not established Inactivation

Activators Retigabine (KCNQ3 alone:  $0.6 \mu M$ ; KCNQ3/KCNQ5:  $1.4 \mu M$ )<sup>2</sup>; XE991,<sup>3</sup> BMS204352 ( $1 \mu M$ )<sup>4</sup>

Gating inhibitors

Blockers Tetraethyammonium (>30 mM),<sup>5</sup> linopiridine (KCNQ3/KCNQ5: 7.7 μM)<sup>2</sup>

Radioligands None

Channel distribution Brain, testis, retina, colon, eye, head, neck

Determines subthreshold excitability of neurons; KCNQ2 and KCNQ3 coassemble to form the M Physiological functions

current in the brain¹ (see "Comments"); KCNQ2 and KCNQ3 proteins are colocalized in a somatodendritic pattern on pyramidal and polymorphic neurons in the human cortex and

hippocampus<sup>7,8</sup>

Mutations and pathophysiology Benign familial neonatal convulsions (EBN2) (e.g., G263V mutation in the pore)9

Pharmacological significance

Comments

Anticonvulsants (activators), cognition enhancers (blockers)<sup>6</sup> The M current is a slowly activating and deactivating potassium conductance that plays a critical

role in determining the subthreshold excitability of neurons as well as the responsiveness to synaptic inputs; the M current was first described in peripheral sympathetic neurons, and differential expression of this conductance produces subtypes of sympathetic neurons with distinct Downloaded from pharmrev.aspetjournals.org by guest on June

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firing patterns; the M current is also expressed in many neurons in the central nervous system



aa, amino acids; chr., chromosome; XE991 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1H-

<sup>1.</sup> Wang HS, Pan Z, Shi W, Brown BS, Wymore RS, Cohen IS, Dixon JE, and McKinnon D (1998) KCNQ2 and KCNQ3 potassium channel subunits: molecular correlates of the M-channel. Science (Wash DC) 282:1890-1893.

<sup>2.</sup> Wickenden AD, Zou A, Wagoner PK, and Jegla T (2001) Characterization of KCNQ5/Q3 potassium channels expressed in mammalian cells. Br. J. Pharmacol 132:381-384 3. Wang HS, Brown BS, McKinnon D, and Cohen IS (2000) Molecular basis for differential sensitivity of KCNQ, and I<sub>Ks</sub> channels to the cognitive enhancer XE991. Mol

Pharmacol 57:1218-1223. 4. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, and Olesen SP (2001) KCNQ4 channel activation by BMS-204352 and retigabine. Neurophar-

macology 40:888-898. 5. Hadley JK, Noda M, Selyanko AA, Wood IC, Abogadie FC, and Brown DA (2000) Differential tetraethylammonium sensitivity of KCNQ1-4 potassium channels. Br J

Pharmacol 129:413-415. 6. Coghlan MJ, Carroll WA, and Gopalakrishnan M (2001) Recent develo pMents in the biology and medicinal chemistry of potassium channel modulators: update from

a decade of progress. J Med Chem 44:1627–1653. 7. Smith JS, Iannotti C, Dargis P, Christian EP, and Aiyar J (2001) Differential expression of KCNQ2 splice variants: implications to M current function during neuronal

develo pMent. J Neurosci 21:1096-1103. 8. Cooper EC, Aldape KD, Abosch A, Barbaro NM, Berger MS, Peacock WS, Jan YN, and Jan LY (2000) Colocalization and coassembly of two human brain M-type potassium channel subunits that are mutated in epilepsy. Proc Natl Acad Sci USA 97:4914-4919.

<sup>9.</sup> Charlier C, Singh NA, Ryan SG, Lewis TB, Reus BE, Leach RJ, and Leppert M (1998) A pore mutation in a novel KQT-like potassium channel gene in an idiopathic epilepsy family. Nat Genet 18:53-55.

### TABLE 27 $K_{\rm v}7.4$ channels

 $K_V 7.4$ Channel name

Description Voltage-gated potassium channel, delayed rectifier

Other names

Human: 695aa, NM\_004700 (transcript variant 1), chr. 1p34, KCNQ4, GeneID: 9132, PMID: Molecular information

Mouse: 724aa, XM\_143960 (predicted), chr. 4 Rat: AF249748 (partial coding sequence)

Associated subunits KCNQ3<sup>2</sup> Functional assays Voltage-clamp Current IK,n

Conductance Not established

Ion selectivity  $K^{+}$ 

Activation  $V_{\rm a} = 10 \text{ mV}$ Inactivation Not established

Activators Retigabine  $(1 \mu M)^3$ ; BMS204352  $(1 \mu M)^3$ 

Gating inhibitors

Blockers Tetraethyammonium (3 mM), <sup>4</sup> linopirdine (14  $\mu$ M), <sup>5</sup> XE991 (5  $\mu$ M), <sup>5</sup> bepridil (9.4  $\mu$ M)<sup>5</sup>

Radioligands

Channel distribution Cochlea (outer hair cells), placenta, vestibular organs (type 1 hair cells), brainstem auditory nuclei

Physiological functions Mediates potassium efflux from outer hair cells<sup>1,6</sup>

Mutations in KCNQ4 cause autosomal dominant nonsyndromic deafness type 2 (DFNA2)<sup>1,6</sup> Mutations and pathophysiology

Pharmacological significance Anticonvulsants (activators)

aa, amino acids; chr., chromosome; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1Hindol-2-one.

- 1. Kubisch C, Schroeder BC, Friedrich T, Lutjohann B, El-Amraoui A, Marlin S, Petit C, and Jentsch TJ (1999) KCNQ4, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness. Cell 96:437-446.
- 2. Schroeder BC, Waldegger S, Fehr S, Bleich M, Warth R, Greger R, and Jentsch TJ (2000) A constitutively open potassium channel formed by KCNQ1 and KCNE3. Nature (Lond) 403:196-199 3. Schroder RL, Jespersen T, Christophersen P, Strobaek D, Jensen BS, and Olesen SP (2001) KCNQ4 channel activation by BMS-204352 and retigabine. Neurophar-
- macology 40:888-898. 4. Hadley JK, Noda M, Selvanko AA, Wood IC, Abogadie FC, and Brown DA (2000) Differential tetraethylammonium sensitivity of KCNQ1-4 potassium channels. Br J
- Pharmacol 129:413-415.
- 5. Sogaard R, Ljungstrom T, Pedersen KA, Olesen SP, and Jensen BS (2001) KCNQ4 channels expressed in mammalian cells: functional characteristics and pharmacology. Am J Physiol 280:C859-C866.
- 6. Kharkovets T, Hardelin JP, Safieddine S, Schweizer M, El-Amraoui A, Petit C, and Jentsch TJ (2000) KCNQ4, a K+ channel mutated in a form of dominant deafness, is expressed in the inner ear and the central auditory pathway. Proc Natl Acad Sci USA 97:4333-4338.

### TABLE 28 $K_V7.5$ channels

Channel name  $K_v7.5$ 

Description Voltage-gated potassium channel, delayed rectifier

Other names

Human: 932aa, NM\_019842, chr. 6q14, KCNQ, 1,5 GeneID: 56479, PMID: 107874161 Molecular information

> Mouse: 933aa, NM\_023872, chr. 1 Rat: 953aa, XM\_237012 (predicted), chr. 9

Associated subunits KCNQ3 Functional assays Voltage-clamp Current M current<sup>1</sup> Conductance Not established

 $K^{+}$ Ion selectivity

Activation  $V_a = 30 \text{ mV}$ Not established Inactivation

Retigabine (KCNQ5/KCNQ3: 1.4  $\mu$ M), BMS204352 (2.4  $\mu$ M) Activator

Gating inhibitors None

Tetraethyammonium (>30 mM), linopiridine (16 μM), linopiridine KCNQ5/KCNQ3 (7.7 μM), Blockers

 $XE991^{3}$ 

Radioligands None

Channel distribution Brain, sympathetic ganglia (splice variant I),<sup>4</sup> skeletal muscle (splice variant III)<sup>4</sup>

Physiological functions Determines subthreshold excitability of neurons Mutations and pathophysiology A number of allelic variants have been identified

Pharmacological significance Anticonvulsants (activators)

aa, amino acids; chr., chromosome; XE991, 10,10-bis(pyridin-4-ylmethyl)anthracen-9-one; BMS204352, 3-(5-chloro-2-methoxy-phenyl)-3-fluoro-6-(trifluoromethyl)-1Hindol-2-one 1. Lerche C, Scherer CR, Seebohm G, Derst C, Wei AD, Busch AE, and Steinmeyer K (2000) Molecular cloning and functional expression of KCNQ5, a potassium channel

- subunit that may contribute to neuronal M-current diversity. J Biol Chem 275:22395-22400 2. Wickenden AD, Zou A, Wagoner PK, and Jegla T (2001) Characterization of KCNQ5/Q3 potassium channels expressed in mammalian cells. Br J Pharmacol
- 3. Dupuis DS, Schroder RL, Jespersen T, Christensen JK, Christophersen P, Jensen BS, and Olesen SP (2002) Activation of KCNQ5 channels stably expressed in HEK293
- cells by BMS-204352. Eur J Pharmacol 437:129-137.

  4. Schroeder BC, Hechenberger M, Weinreich F, Kubisch C, and Jentsch TJ (2000) KCNQ5, a novel potassium channel broadly expressed in brain, mediates M-type

currents. J Biol Chem 275:24089-24095.



# Spet

# TABLE 29 $K_v 8.1$ channels

 $\begin{array}{lll} \text{Channel name} & & K_V 8.1^{1-3} \\ \text{Description} & & \text{Modifier/silencer} \\ \text{Other names} & & K_V 2.3, \, \text{HNKA} \end{array}$ 

Molecular information Human: 500aa, NM\_014379, chr. 8q22.3-24.1, KCNV1, GeneID: 27012, PMID: 6708331

Mouse: 503aa, NM\_026200, chr. 15 Rat: 503aa, NM\_021697, chr. 7q31 Coassembles with  $K_{\rm V}2$  family channels

Functional assays Voltage-clamp Current None established

Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Associated subunits

Channel distribution Infant brain, adult brain (layers II, IV, and VI of the cerebral cortex, hippocampus, CA1-CA4

pyramidal cell layer, granule cells of the dentate gyrus, granule cell layer, Purkinje cell layer of

the cerebellum), kidney

Physiological functions Regulation of membrane potential and action potential frequency by modulation of delayed rectifier

potassium current; modulates the activity of  $K_{\rm V}2.1$  and  $K_{\rm V}2.2$  channels by changing kinetics and

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levels of expression and by shifting the half-inactivation potential to more polarized values

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments  $K_V 8.1$  has no function on its own, but it has important modulatory actions on  $K_V 2$  channels

aa, amino acids; chr., chromosome.

1. Hugnot JP, Salinas M, Lesage F, Guillemare E, de Weille J, Heurteaux C, Mattei MG, and Lazdunski M (1996) Kv8.1, a new neuronal potassium channel subunit with specific inhibitory properties towards Shab and Shaw channels. EMBO J 15:3322–3331.

2. Salinas M, de Weille J, Guillemare E, Lazdunski M, and Hugnot JP (1997) Modes of regulation of Shab K<sup>+</sup> channel activity by the Kv8.1 subunit. J Biol Chem 272:8774–8780.

3. Chiara MD, Monje F, Castellano A, and Lopez-Barneo J (1999) A small domain in the N terminus of the regulatory  $\alpha$ -subunit Kv2.3 modulates Kv2.1 potassium channel gating. J Neurosci 19:6865–6873.

# TABLE 30 $K_{v}8.2$ channels

Channel name  $K_V 8.2$ 

Description Modifier/silencer

Other names Kv11.1<sup>1</sup>

Molecular information Human: 545aa, NM\_133497, chr. 9p24.2, KCNV2, GeneID: 169522, PMID: 12060745<sup>1</sup>

Mouse: 562aa, NM\_183179, chr. 19 Rat: 561 aa, XM\_220024 (predicted), chr. 1 Coassembles with K.-2 family channels

Associated subunits  $Coassembles with K_V2 family channels Functional assays$  Voltage-clamp

Current None established
Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Lung, liver, kidney, pancreas, spleen, thymus, prostate, testis, ovary, colon<sup>1</sup>

Physiological functions Regulation of membrane potential and action potential frequency by modulation of delayed rectifier

potassium currents; modulates the activity of  $K_{\rm V}2.1$  channels by causing small changes in

activation threshold and kinetics and in C-type inactivation<sup>1</sup>

Mutations and pathophysiology Not established Pharmacological significance Not established

Comments  $K_{V}8.2$  has no function on its own, but it has important modulatory actions on  $K_{V}2$  channels

aa, amino acids; chr., chromosome.

Ottschytsch N, Raes A, Van Hoorick D, and Snyders DJ (2002) Obligatory heterotetramerization of three previously uncharacterized Kv channel-subunits identified in the human genome. Proc Natl Acad Sci USA 99:7986-7991.

# TABLE 31 $K_{\rm V}9.1$ channels

 $\begin{array}{lll} \text{Channel name} & & \text{K}_{\text{V}}9.1^{1-4} \\ \text{Description} & & \text{Modifier/silencer} \\ \text{Other names} & & \text{None} \end{array}$ 

Molecular information Human: 526 aa, NM\_002251, chr. 20q12, KCNS1, GeneID: 3787, PMID: 10484328<sup>3</sup>

Mouse: 497 aa, NM\_008435, chr. 2 Rat: 497 aa, NM\_053954, chr. 3q42 Coassembles with  $K_{\rm V}2$  family channels

Functional assays Voltage-clamp
Current None established
Conductance Not functional on

Conductance Not functional on its own Ion selectivity Not functional on its own Activation Not functional on its own Inactivation Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Associated subunits

Channel distribution Infant brain, adult brain (frontal cortex), lens epithelium, melanocytes (in mouse brain, the distribution of  $K_V 9.1$  is similar to  $K_V 9.2$ , with highest expression levels in the main olfactory bulb,

cerebral cortex, hippocampal formation, habenula, basolateral amygdaloid nuclei, and cerebellum;  $K_V 9.1$  and  $K_V 9.2$  are colocalized with  $K_V 2.1$  and/or  $K_V 2.2$   $\alpha$  subunits in several regions)

Physiological functions Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium current; modulates the activity of  $K_V2.1$  and  $K_V2.2$   $\alpha$  subunits by changing kinetics and

levels of expression and by shifting the half-inactivation potential to more polarised values;  $K_{\rm v}9.1$  enhances the single-channel conductance of  $K_{\rm v}2.1$ 

Mutations and pathophysiology Pharmacological significance Not established Not established

Comments The human  $K_{v}9$ 

The human  $K_{\rm V}9.1$  gene is composed of a minimum of 5 exons, with at least 2 alternatively spliced exons in the 5'-untranslated region<sup>3</sup>

aa, amino acids; chr., chromosome.

1. Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian Shab K<sup>+</sup> channels. J Biol Chem **272**:24371–24379.

2. Stocker M and Kerschensteiner D (1998) Cloning and tissue distribution of two new potassium channel α-subunits from rat brain. Biochem Biophys Res Commun **248**:927–934.

3. Shepard AR and Rae JL (1999) Electrically silent potassium channel subunits from human lens epithelium. Am J Physiol 277:C412-C424.

4. Richardson FC and Kaczmarek LK (2000) Modification of delayed rectifier potassium currents by the Kv9.1 potassium channel subunit. Hear Res 147:21-30.

### TABLE 32 K<sub>v</sub>9.2 channels

 $\begin{array}{lll} \text{Channel name} & & \text{$K_{V}9.2^{1,2}$} \\ \text{Description} & & \text{Modifier/silencer} \\ \text{Other names} & & \text{None} \\ \end{array}$ 

Molecular information Human: 477aa, NM 020697, chr. 8q22, KCNS2, GeneID: 3788, PMID: 9305895<sup>1</sup>

Mouse: 477aa, NM\_181317, chr. 15 Rat: 477 aa, NM\_023966, chr. 7q22

Current None established
Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own
Inactivation Not functional on its own
Not functional on its own

Activators None
Gating inhibitors None
Blockers None
Radioligands None

Channel distribution Infant and adult brain, retina, spinal cord (in mouse brain, the distribution of  $K_{\rm V}9.2$  is similar to

 $K_V9.1$ , with highest expression levels in the main olfactory bulb, cerebral cortex, hippocampal formation, habenula, basolateral amygdaloid nuclei, and cerebellum;  $K_V9.1$  and  $K_V9.2$  are colocalized with  $K_V2.1$  and/or  $K_V2.2$   $\alpha$  subunits in several regions; also found in the retina, spinal

cord, and pulmonary artery)

Physiological functions Regulation of membrane potential and action potential frequency by modulation of delayed rectifier potassium current; modulates the activity of  $K_v2.1$  and  $K_v2.2$   $\alpha$  subunits by changing kinetics and

levels of expression and by shifting the half-inactivation potential to more polarized values;  $K_v9.1$  enhances the single-channel conductance of  $K_v2.1$ 

Mutations and pathophysiology Not established Pharmacological significance Not established

aa, amino acids; chr., chromosome.

<sup>1.</sup> Salinas M, Duprat F, Heurteaux C, Hugnot JP, and Lazdunski M (1997) New modulatory α subunits for mammalian Shab K<sup>+</sup> channels. J Biol Chem 272:24371–24379.

<sup>2.</sup> Davies AR and Kozlowski RZ (2001) Kv channel subunit expression in rat pulmonary arteries. Lung 179:147–161.
3. Banfi S, Borsani G, Rossi E, Bernard L, Guffanti A, Rubboli F, Marchitiello A, Giglio S, Coluccia E, Zollo M, et al. (1996) Identification and mapping of human cDNAs homologous to Drosophila mutant genes through EST database searching. Nat Genet 13:167–174.

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# TABLE 33 $K_{v}9.3$ channels

Other names None

Associated subunits

Molecular information Human: 491aa, NM\_023966, NM\_002252, chr. 2p24, KCNS3 (see 'Comments'), GeneID: 3790,

PMID: 93624761

Mouse: 491aa, NM\_173417, chr. 12 Rat: 491aa, NM\_031778, chr. 6q14 Coassembles with  $K_{\rm V}2$  family channels

Functional assays Voltage-clamp

Current K<sub>V</sub>9.3/K<sub>V</sub>2.1 and ATP-dependent delayed rectifier channel in oxygen-sensitive pulmonary myocytes

Conductance Not functional on its own
Ion selectivity Not functional on its own
Activation Not functional on its own

Inactivation  $K_V 9.3/K_V 2.1$  heteromers inactivate in a fast and complete fashion from intermediate closed states

but in a slow and incomplete manner from open states<sup>4</sup>

Activators None Gating inhibitors None

Blockers Hypoxia blocks  $K_V 9.3/K_V 2.1$  channels<sup>5</sup>

Radioligands None

Channel distribution Brain, breast, colon, eye, lens, heart, kidney, muscle, lung, testis, skin, stomach, uterus<sup>6</sup>; also found

in lens epithelium<sup>3</sup>

Physiological functions Regulation of membrane potential in pulmonary artery myocytes

Mutations and pathophysiology Not established

Pharmacological significance Pulmonary artery hypertension

Comments The human  $K_V 9.3$  gene is intronless across the coding region 3'-UTR and all of the analysed 5'-UTR

aa, amino acids; chr., chromosome; UTR, untranslated region.

1. Patel AJ, Lazdunski M, and Honore E (1997) Kv2.1/Kv9.3, a novel ATP-dependent delayed-rectifier K<sup>+</sup> channel in oxygen-sensitive pulmonary artery myocytes. EMBO J 16:6615–6625.

2. Stocker M and Kerschensteiner D (1998) Cloning and tissue distribution of two new potassium channel  $\alpha$ -subunits from rat brain. Biochem Biophys Res Commun 248:927–934.

3. Shepard AR and Raem JL (1999) Electrically silent potassium channel subunits from human lens epithelium. Am J Physiol 277, C412–C424.

4. Kerschensteiner D and Stocker M (1999) Heteromeric assembly of Kv2.1 with Kv9.3: effect on the state dependence of inactivation. Biophys J 77:248–257.

5. Hulme JT, Coppock EA, Felipe A, Martens JR, and Tamkun MM (1999) Oxygen sensitivity of cloned voltage-gated K<sup>+</sup> channels expressed in the pulmonary vasculature. Circ Res 85:489–497.

 $6. \ UniGene Cluster \ Hs 0.47584; \ OMIM \ no. \ 603888. \\$ 



### TABLE 34 $K_{\rm V}10.1$ channels

Channel name K<sub>v</sub>10.1

Associated subunits

Description Voltage-gated potassium channel, delayed rectifier eag1a, eag1b, KCNH1a, KCNH1b, ether-à-go-go<sup>1-4</sup> Other names

Human: 989aa, NM\_172362, chr. 1q32-41, KCNH1 (see "Comments"), GeneID: 3756. Molecular information

PMID: 8159766<sup>2</sup>

Mouse: 989aa, NM\_010600, chr. 1 Rat: 962aa, NM\_031742, chr. 13q27

Hyperkinetic (Hk),<sup>5</sup> CaM,<sup>6</sup> Slob,<sup>7</sup> epsin,<sup>8</sup> KCR1 (K channel regulator)<sup>9</sup>

Voltage-clamp Functional assays Current Delayed rectifier Conductance Not established K<sup>+</sup> and Ca,<sup>2+ 10</sup> variable Cs<sup>+</sup> Ion selectivity

Extracellular Mg<sup>2+</sup> and other divalent cations slow activation in a dose- and voltage-dependent Activation

manner, based on their enthalpy of hydration<sup>11</sup>; low external pH also slows activation

Inactivation Not established

Activators Hyperpolarization slows down the kinetics of activation; depolarization accelerates the kinetics of

activation3

None Gating inhibitors

Quinidine (1.4 µM),12 calcium/calmodulin (480 nM)6,13 Blockers

Radioligands None

Channel distribution Brain (amygdala, caudate nucleus, cerebral cortex, cerebellum, putamen, hippocampus, frontal lobe, occipital lobe, temporal lobe, subthalamic nucleus; not in substantia nigra, thalamus, or medulla

oblongata), myoblasts, skeletal muscle (ESTs, but not detected by Northern), melanoma cells, ectopic expression in cancer cell lines and many tumor cells from different tissues, spiral ligament in rat<sup>14–16</sup> Role in controlling the cell cycle and/or cell proliferation<sup>17,18</sup>; eag-1 is thought to encode the

noninactivating delayed rectifier potassium channel  $K_{\rm NI}$  that is activated at the onset of human myoblast differentiation<sup>4</sup>

Mutations and pathophysiology Pharmacological significance Comments

Physiological functions

K<sub>v</sub>10.1 has been associated with human cervical carcinoma<sup>21</sup>

K<sub>v</sub>10.1 blockers might have use in cancer therapy

This channel has a GFG (rather than the common GYG) potassium channel signature sequence, a PAS domain in the distal part of the cytosolic N terminus, a cNBD domain in the proximal portion of the C terminus, a C-terminal assembly domain (CAD), a CaM-binding domain, a bNLS domain in the C terminus, and a C-terminal domain required for assembly 19; the TCC domain at the C-terminal end of K, 10 and K, 11 confers specificity for multimer formation, allowing K, 10.1/ K<sub>v</sub>10.2 heteromerization and K<sub>v</sub>11.1 homomerization but not K<sub>v</sub>10.x/K<sub>v</sub>11.1 heteromerization<sup>22</sup>; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC have been found to be linked to genetic channelopathies; conductance properties have been shown to change with the cell cycle<sup>20</sup>

aa, amino acids; chr., chromosome; CaM, calmodulin; TCC, tetramerizing coiled-coiled; EST, expressed sequence tag.

- 1. Warmke J, Drysdale R, and Ganetzky B (1991) A distinct potassium channel polypeptide encoded by the Drosophila eag locus. Science (Wash DC) 252:1560-1562.
- 2. Warmke JW and Ganetzky B (1994) A family of potassium channel genes related to eag in Drosophila and mammals. Proc Natl Acad Sci USA 91:3438-3442. 3. Ludwig J, Terlau H, Wunder F, Bruggemann A, Pardo LA, Marquardt A, Stuhmer W, and Pongs O (1994) Functional expression of a rat homologue of the voltage gated
- ether à go-go potassium channel reveals differences in selectivity and activation kinetics between the Drosophila channel and its mammalian counterpart. EMBO J 13:4451-4458.
- 4. Occhiodoro T, Bernheim L, Liu JH, Bijlenga P, Sinnreich M, Bader CR, and Fischer-Lougheed J (1998) Cloning of a human ether-à-go-go potassium channel expressed in myoblasts at the onset of fusion. FEBS Lett 434:177-182.
- 5. Wilson GF, Wang Z, Chouinard SW, Griffith LC, and Ganetzky B (1998) Interaction of the K channel β subunit, Hyperkinetic, with eag family members. J Biol Chem **273:**6389-6394.
  - Schonherr R, Lober K, and Heinemann SH (2000) Inhibition of human ether à go-go potassium channels by Ca<sup>2+</sup>/calmodulin. EMBO J 19:3263-3271.
- 7. Schopperle WM, Holmqvist MH, Zhou Y, Wang J, Wang Z, Griffith LC, Keselman I, Kusinitz F, Dagan D, and Levitan IB (1998) Slob, a novel protein that interacts with the Slowpoke calcium-dependent potassium channel. Neuron 20:565-573.
- 8. Piros ET, Shen L, and Huang XY (1999) Purification of an EH domain-binding protein from rat brain that modulates the gating of the rat ether-à-go-go channel. J Biol Chem 274:33677-33683.
- 9. Hoshi N, Takahashi H, Shahidullah M, Yokoyama S, and Higashida H (1998) KCR1, a membrane protein that facilitates functional expression of non-inactivating K<sup>+</sup> currents associates with rat EAG voltage-dependent K<sup>+</sup> channels. *J Biol Chem* **273**:23080–23085.

  10. Bruggemann A, Pardo LA, Stuhmer W, and Pongs O (1993) *Ether-à-go-go* encodes a voltage-gated channel permeable to K<sup>+</sup> and Ca<sup>2+</sup> and modulated by cAMP. *Nature*
- 11. Terlau H, Ludwig J, Steffan R, Pongs O, Stuhmer W, and Heinemann SH (1996) Extracellular Mg<sup>2+</sup> regulates activation of rat eag potassium channel. Pflueg Arch
- Eur J Physiol 432:301-312.
- 12. Schonherr R, Gessner G, Lober K, and Heinemann SH (2002) Functional distinction of human EAG1 and EAG2 potassium channels. FEBS Lett 514:204-208. 13. Stansfeld CE, Roper J, Ludwig J, Weseloh RM, Marsh SJ, Brown DA, and Pongs O (1996) Elevation of intracellular calcium by muscarinic receptor activation induces a block of voltage-activated rat ether-à-go-go channels in a stably transfected cell line. Proc Natl Acad Sci USA 93:9910-9914.
- 14. Lecain E, Sauvaget E, Crisanti P, Van Den Abbeele T, and Huy PT (1999) Potassium channel ether à go-go mRNA expression in the spiral ligament of the rat. Hear Res 133:133-138.
- 15. Meyer R, Schonherr R, Gavrilova-Ruch O, Wohlrab W, and Heinemann SH (1999) Identification of ether à go-go and calcium-activated potassium channels in human melanoma cells. J Membr Biol 171:107-115.
- 16. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K+ channels in brain. J Neurosci **21:**4609-4624
- $17. \ Pardo\ LA,\ del\ Camino\ D,\ Sanchez\ A,\ Alves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ W\ (1999)\ Oncogenic\ potential\ of\ EAG\ K^+\ channels.\ EMBO\ J\ 18:5540-5547\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ W\ (1999)\ Oncogenic\ potential\ of\ EAG\ K^+\ channels.\ EMBO\ J\ 18:5540-5547\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ W\ (1999)\ Oncogenic\ potential\ of\ EAG\ K^+\ channels.\ EMBO\ J\ 18:5540-5547\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ W\ (1999)\ Oncogenic\ potential\ of\ EAG\ K^+\ channels.\ EMBO\ J\ 18:5540-5547\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ W\ (1999)\ Oncogenic\ potential\ of\ EAG\ K^+\ channels.\ EMBO\ J\ 18:5540-5547\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ B,\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ B,\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ B,\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ and\ Stuhmer\ B,\ Adves\ F,\ Bruggemann\ A,\ Beckh\ S,\ Adves\ F,\ Bruggemann\ A,\ Beckh\ Bruggemann\ A,\ Bruggemann\ A,\ Beckh\ Bruggemann\ A,\ Bruggemann$ 18. Camacho J, Sanchez A, Stuhmer W, and Pardo LA (2000) Cytoskeletal interactions determine the electrophysiological properties of human EAG potassium channels. Pflueg Arch Eur J Physiol 441:167-174.
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Channel name

 $K_{\rm v}10.2$  channels



 $K_{v}10.2$ 

Description Outward-rectifying, noninactivating voltage-dependent K<sup>+</sup> currents<sup>3-5</sup>

 $eag2^{1-5}$ Other names

Human: 987aa, NM\_139318 (transcript variant 1), chr. 14q23.1, KCNH5 (see "Comments"), Molecular information

> GeneID: 27133, PMID: 97384732 Mouse: 988aa, NM\_172805, chr. 12 Rat: 988aa, NM\_133610, chr. 6q24

Associated subunits Hyperkinetic (Hk), <sup>6</sup> CaM, Slob, KCR1 (potassium channel regulator)

Functional assays Voltage-clamp Current Outward-rectifying Conductance Not established

Ion selectivity K

Activates at -100 mV (rat)3 Activation

Inactivation Noninactivating

Activators None Gating inhibitors None

Blockers Quinidine (152 μM),<sup>5</sup> intracellular calcium (nanomolar)<sup>4</sup>

Radioligands

Channel distribution Brain (layer IV of the cerebral cortex; thalamus, inferior colliculus, olfactory bulb, and certain

brainstem nuclei)<sup>3,4</sup>

Physiological functions Not established Mutations and pathophysiology Pharmacological significance

Not established Not established

Comments This channel has a GFG (rather than the common GYG) potassium channel signature sequence, a

PAS domain in the distal part of the cytosolic N terminus, a cNBD domain in the proximal portion of the C terminus, a C-terminal assembly domain (CAD), a CaM-binding domain, a bNLS domain in the C terminus, and a C-terminal domain is required for assembly<sup>7</sup>; the TCC domain at the C-terminal end of K<sub>v</sub>10 and K<sub>v</sub>11 confers specificity for multimer formation, allowing K<sub>v</sub>10.1/ K<sub>v</sub>10.2 heteromerization and K<sub>v</sub>11 homomerization but not K<sub>v</sub>10.x/K<sub>v</sub>11.x heteromerization<sup>8</sup>; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC

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have been found to be linked to genetic channelopathies

aa, amino acids; chr., chromosome; CaM, calmodulin; TCC, tetramerizing coiled-coiled.

<sup>1.</sup> Shi W, Wang HS, Pan Z, Wymore RS, Cohen IS, McKinnon D, and Dixon JE (1998) Cloning of a mammalian elk potassium channel gene and EAG mRNA distribution in rat sympathetic ganglia. J Physiol 511:675-682.

<sup>2.</sup> Occhiodoro T, Bernheim L, Liu JH, Bijlenga P, Sinnreich M, Bader CR, and Fischer-Lougheed J (1998) Cloning of a human ether-à-go-go potassium channel expressed in myoblasts at the onset of fusion. FEBS Lett 434:177-182. 3. Saganich MJ, Vega-Saenz de Miera E, Nadal MS, Baker H, Coetzee WA, and Rudy B (1999) Cloning of components of a novel subthreshold-activating K+ channel with

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<sup>7.</sup> Ludwig J, Owen D, and Pongs O (1997) Carboxy-terminal domain mediates assembly of the voltage-gated rat ether-à-go-go potassium channel. EMBO J 16:6337-6345. 8. Jenke M, Sanchez A, Monje F, Stuhmer W, Weseloh RM, and Pardo LA (2003) C-terminal domains implicated in the functional surface expression of potassium channels. EMBO J 22:395-403.

### TABLE 36 $K_{\rm V}11.1$ channels

 $K_v11.1$ Channel name

Description Voltage-gated potassium channel with inwardly rectifying properties

Other names Human ether-à-go-go-related gene, HERG, erg1, Hergb<sup>1-8</sup>

Human: 1159aa, NM\_000238 (transcript variant 1), chr. 7q35-36, KCNH2, GeneID: 3757, Molecular information

PMID: 81597661

Mouse: 1162aa, NM\_013569, chr. 5 Rat: 1163aa, NM\_053949, chr. 4q11 minK,9,25 possibly MiRP1 (KCNE2)10

Functional assays Voltage-clamp

Cardiac  $\bar{I_{\rm Kr}}$  current  $^{3,26}$ Current

Conductance 2pS (in physiological [K] $_{o}$ ), 10pS (100 mM [K] $_{o}$ )<sup>11</sup>

Ion selectivity K

Associated subunits

Comments

Activation Activation at currents more positive than -50 mV<sup>3,26</sup>

Inactivation Exhibits C-type inactivation<sup>4</sup>; inward rectification arises from a rapid and voltage-dependent

inactivation process that reduces conductance at positive voltages<sup>3,26,27</sup>

Activators None Gating inhibitors None

Astemizole (1 nM), <sup>13</sup> BeKM-1 (3 nM), <sup>14</sup> ergtoxin (12 nM), <sup>15</sup> sertindole (3 nM), dofetilide Blockers

> (15–35 nM), <sup>16</sup> cisapride (6–40 nM), pimozide (18 nM), terfenadine (56 nM), halofantrine (200 nM), BRL32872 (240 nM), E-4031 (7.7 nM), CT haloperidol (1  $\mu$ M), imipramine (3  $\mu$ M), cocaine (5  $\mu$ M),

ketoconazole

Radioligands

Channel distribution Heart, leiomyosarcoma, hippocampus, neuroblastoma, blood cells, brain, kidney, liver, lung, ovary,

pancreas, testis, prostate, small intestine, tonsil, uterus, microglia

HERG proteins form cardiac  $I_{Kr}$  channels  $^{3,26}$ ; in the heart, HERG channels produce a resurgent Physiological functions

current during repolarization<sup>20</sup> due to the recovery from C-type inactivation<sup>4</sup> and a slow deactivation due to an interaction with an N-terminal domain (AA2-16) and the internal mouth of the pore<sup>1,22</sup>; HERG contains a tetramerization domain called NAB and a structurally defined PAS domain in distinct regions of the N terminus<sup>17</sup>; HERG forms a complex with MiRP1,<sup>10</sup> but it is as yet unclear whether MiRP1 forms a stable part of the channel itself or is otherwise involved in

regulation of HERG expression or stability<sup>23</sup>

Mutations of this gene cause the autosomal dominant long QT syndrome 2 due to gating defects<sup>28</sup> Mutations and pathophysiology and trafficking abnormalities<sup>29–33</sup> and a prolonged QT interval on the electrocardiogram; syncope,

sudden cardiac death, ventricular fibrillation, and torsades de pointes are also implicated in acquired long QT syndrome; mutations in MiRP1 are the cause of long QT syndrome 6 and are

also found in many tumors  $^{18,19}$ 

Proarrhythmic potential (QT prolongation) of histamine H<sub>1</sub> receptor antagonists, antipsychotics, and Pharmacological significance tricyclic antidepressants that leads to torsades de points in some individuals (acquired long QT

syndrome)

A shorter isoform encoded by an alternative transcript (1b) of K<sub>v</sub>11.1<sup>5,7</sup> or a truncated isoform<sup>6</sup> can coassemble with and modulate the behavior of full-length HERG and Merg1, the mouse ortholog; the TCC domain at the C-terminal end of K<sub>v</sub>10 and K<sub>v</sub>11 confers specificity for multimer formation, allowing K<sub>v</sub>10.1/K<sub>v</sub>10.2 heteromerization and K<sub>v</sub>11 homomerization, but not K<sub>v</sub>10.x/ K, 11.x heteromerization<sup>24</sup>; this C-terminal TCC domain has been identified in many other channels, and mutations of the TCC are found to be linked to genetic channel opathies; C terminus

interacts with Golgi matrix protein GM130<sup>34</sup>

aa, amino acids; chr., chromosome; MiRP1, MinK-related peptide 1; TCC, tetramerizing coiled-coiled; E-4031, N-[4-[[1-[2-(6-methyl-2-pyridinyl)ethyl]-4-piperidinyl]carbonyl]phenyl]methanesulfonamide dihydrochloride.

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<sup>13.</sup> Suessbrich H, Waldegger S, Lang F, and Busch AE (1996) Blockade of HERG channels expressed in Xenopus oocytes by the histamine receptor antagonists terfenadine and astemizole. FEBS Lett 385:77-80.

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### TABLE 37 K<sub>v</sub>11.2 channels

Channelname K<sub>v</sub>11.2

Description Voltage-gated potassium channel

Other names  $erg2^{1,2}$ 

Molecular information Human: 994aa, NM\_030779 (transcript variant 1) chr. 17q23.3, KCNH6, GeneID: 81033,

PMID: 10414305<sup>6</sup>

Rat: 950aa, NM\_053937, chr. 10q32.1

Associated subunits
Functional assays
Voltage-clamp
Current
Not established
Conductance
Not established
Ion selectivity
Not established
Activation
Not established
Inactivation
Not established
Not established

Activators None
Gating inhibitors None
Blockers Sipatrigine
Radioligands None

Channel distribution Brain,<sup>2</sup> uterus, leiomyosarcoma, hippocampus, neuroblastoma, lactotrophs,<sup>3</sup> GH3/B6 cells, rat

pituitary<sup>4</sup>

Physiological functions
Mutations and pathophysiology
Pharmacological significance
Not established
Not established

Comments  $K_V11.1, K_V11.2, \text{ and } K_V11.3 \text{ can form heteromultimers}^5$ 

aa, amino acids; chr., chromosome.

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- 3. Shi W, Wymore RS, Wang HS, Pan Z, Cohen IS, McKinnon D, and Dixon JE (1997) Identification of two nervous system-specific members of the erg potassium channel gene family. J Neurosci 17:9423–9432.
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  - 6. Ganetzky B, Robertson GA, Wilson GF, Trudeau MC, and Titus SA (1999) The eag family of K+ channels in Drosophila and mammals. Ann NY Acad Sci 868:356-369.

### TABLE 38 $K_{\rm V}11.3$ channels

Channel name  $K_{v}11.3$ 

Description Voltage-gated potassium channel

 $erg3^{1-}$ Other names

Human: 1196aa, NM\_033272 (transcript variant 1), chr. 2q24.2, KCNH7, GeneID: 90134, Molecular information

PMID: 10414305<sup>9</sup>

Mouse: 1195aa, NM\_133207, chr. 2 Rat: 1195aa, NM\_131912, chr. 3q21

Associated subunits See "Comments" Functional assays Voltage-clamp Current Not established Conductance Not established

Ion selectivity

Activation Activated at −50 mV<sup>2</sup> (see "Comments")

Inactivation Not established

Activators None Gating inhibitors None

Blockers Sertindole (43 nM)<sup>2</sup> and pimozide (103 nM)<sup>2</sup>

Radioligands

Brain, sympathetic ganglia, CA pyramidal neurons, 4 lactotrophs, 5 GH3/B6 cells, rat pituitary 6 Channel distribution

Physiological functions Not established Not established Mutations and pathophysiology Pharmacological significance Not established

Comments Thyrotropin-releasing hormone reduces K<sub>v</sub>11.3 currents and shifts the voltage dependence of

activation by 6 mV<sup>7</sup>; K<sub>v</sub>11.1, K<sub>v</sub>11.2, and K<sub>v</sub>11.3 can form heteromultimers<sup>8</sup>

aa, amino acids; chr., chromosome

1. Shi W, Wymore RS, Wang HS, Pan Z, Cohen IS, McKinnon D, and Dixon JE (1997) Identification of two nervous system-specific members of the erg potassium channel gene family. J Neurosci 17:9423-9432.

2. Kang J, Chen XL, and Rampe D (2001) The antipsychotic drugs sertindole and pimozide block erg3, a human brain K+ channel. Biochem Biophys Res Commun

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4. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K<sup>+</sup> channels in brain. J Neurosci 21:4609-4624.

5. Schafer R, Wulfsen I, Behrens S, Weinsberg F, Bauer CK, and Schwarz JR (1999) The erg-like potassium current in rat lactotrophs. J Physiol 518:401-416.

6. Wulfsen I, Hauber HP, Schiemann D, Bauer CK, and Schwarz JR (2000) Expression of mRNA for voltage-dependent and inward-rectifying K channels in GH3/B6 cells and rat pituitary. J. Neuroendocrinol 12:263-272. 7. Schledermann W, Wulfsen I, Schwarz JR, and Bauer CK (2001) Modulation of rat erg1, erg2, erg3 and HERG K+ currents by thyrotropin-releasing hormone in anterior

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9. Ganetzky B, Robertson GA, Wilson GF, Trudeau MC, and Titus SA (1999) The eag family of K+ channels in Drosophila and mammals. Ann NY Acad Sci 868:356-369.

### TABLE 39 $K_{\rm v}$ 12.1 channels

Channel name  $K_{v}12.1$ 

Slowly activating and deactivating voltage-gated potassium channel<sup>1</sup> Description

Other names elk1.1 elk32

Human: 1107aa, NM\_144633, chr. 3p24.3, KCNH8, GeneID: 131096, PMID: 12890647<sup>3</sup> Molecular information

Mouse: 1102aa, NM\_001031811, chr. 17

Rat: 1102aa, NM\_145095, chr. 9q11 (see "Comments")

Associated subunits Not established Functional assays Voltage-clamp None identified Current Conductance Not established

Ion selectivity  $K^{\dagger}$ 

Not established Activation Inactivation Not established

None Activators Gating inhibitors None  $Ba^{2+1}$ Blockers Radioligands

Sympathetic ganglia, testis, brain, colon, lung, uterus, pre-B cell leukemia (ESTs)<sup>1,2</sup> Channel distribution

Physiological functions Not established Mutations and pathophysiology Not established Pharmacological significance Not established

There is a light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domain in the N and C Comments terminus, respectively.

aa, amino acids; chr., chromosome.

<sup>1.</sup> Shi W, Wang HS, Pan Z, Wymore RS, Cohen IS, McKinnon D, and Dixon JE (1998) Cloning of a mammalian elk potassium channel gene and EAG mRNA distribution in rat sympathetic ganglia J Physiol 511:675-682.

<sup>2.</sup> Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat ether-à-go-go-like K<sup>+</sup> channel genes. J Physiol 513:647–654. 3. Zou A, Lin Z, Humble M, Creech CD, Wagoner PK, Krafte D, Jegla TJ, and Wickenden AD (2003) Distribution and functional properties of human KCNH8 (Elk1) potassium channels. Am J Physiol Cell Physiol 285:C1356-C1366.

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TABLE 40  $K_v$ 12.2 channels

Channel name K<sub>V</sub>12.2

Description Voltage-gated potassium channel

Other names BEC1, Elk2

Molecular information Human: 1083aa, NM\_012284, chr. 12q13, KCNH3, GeneID: 23416, PMID: 104551801

Mouse: 1095aa, NM\_010601, chr. 15 Rat: 1087aa, NM\_017108, 7q36

Associated subunits

Functional assays

Current

Conductance

Not established

Not established

Ion selectivity K<sup>+</sup>

Activation Not established

InactivationFast1,2ActivatorsNoneGating inhibitorsNoneBlockersNoneRadioligandsNone

Channel distribution Infant brain, lung (small cell carcinoma), eye (retinoblastoma), sciatic nerve, cortex, amygdala,

hippocampus (mainly in CA1 and CA3 pyramidal cell body layers and in the granule cell layers of

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the dentate gyrus); in the striatal regions, including the putamen and caudate nucleus,

lymphocytes, leukemias, and NG108-15 cell line<sup>1-5</sup>

Physiological functions
Mutations and pathophysiology
Pharmacological significance
Not established
Not established

Comments There is a light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domain in the N and C

terminus, respectively.

aa, amino acids; chr., chromosome.

1. Miyake A, Mochizuki S, Yokoi H, Kohda M, and Furuichi K (1999) New  $ether-\grave{a}-go-go$  K $^+$  channel family members localized in human telencephalon. J Biol Chem 274:25018–25025.

2. Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat ether-à-go-go-like K+ channel genes. J Physiol 513:647-654.

3. Meves H, Schwarz JR, and Wulfsen I (1999) Separation of M-like current and ERG current in NG108-15 cells. Br J Pharmacol 127:1213-1223.

4. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding subthreshold-operating voltage-gated K<sup>+</sup> channels in brain. J Neurosci 21:4609–4624.

5. Smith GA, Tsui HW, Newell EW, Jiang X, Zhu XP, Tsui FW, and Schlichter LC (2002) Functional up-regulation of HERG K<sup>+</sup> channels in neoplastic hematopoietic cells. J Biol Chem 277:18528–18534.

# TABLE 41 $K_V 12.3$ channels

Channel name K<sub>V</sub>12.3

Description Slowly activating voltage-gated potassium channel

Other names BEC2,<sup>1</sup> elk1<sup>2</sup>

Molecular information Human: 1017aa, NM\_012285, KCNH4, chr. 17q21.2, GeneID: 23415, PMID: 10455180<sup>1</sup>

Rat: 1017aa, NM\_053630, chr. 10q32.1 (see "Comments")

Associated subunits None

Functional assays Voltage-clamp
Current Not established
Conductance Not established

Ion Selectivity K<sup>+</sup>

Activation Threshold for activation is 90 mV<sup>2</sup>

Inactivation Not established

Channel distribution Brain (telencephalon), 1,3 neuroblastoma, esophagus, oligodendroglioma, lung, primary B-cell

neoplasia, cerebellum, pituitary gland<sup>4</sup>

Physiological functions
Mutations and pathophysiology
Pharmacological significance
Not established
Not established

Comments There are light oxygen voltage (LOV) and cyclic nucleotide binding (CNB) domains in the N and C

terminus, respectively.

aa, amino acids; chr., chromosome.

<sup>1.</sup> Miyake A, Mochizuki S, Yokoi H, Kohda M, and Furuichi K (1999) New ether-à-go-go K<sup>+</sup> channel family members localized inhuman telencephalon. J Biol Chem 274:25018–25025.

<sup>2.</sup> Engeland B, Neu A, Ludwig J, Roeper J, and Pongs O (1998) Cloning and functional expression of rat ether-à-go-go-like K<sup>+</sup> channel genes. J Physiol **513**:647–654.

3. Saganich MJ, Machado E, and Rudy B (2001) Differential expression of genes encoding sub threshold-operating voltage-gated K<sup>+</sup> channels in brain. J Neurosci **21**:4609–4624.

<sup>4.</sup> Wulfsen I, Hauber HP, Schiemann D, Bauer CK, and Schwarz JR (2000) Expression of mRNA for voltage-dependent and inward-rectifying K channels in GH3/B6 cells and rat pituitary. J Neuroendocrinol 12:263–272.